

Genomically matched therapy in advanced solid tumors: the randomized phase 2 ROME trial

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A list of authors and their affiliations appears at the end of the paper

Despite recent advancements demonstrating the potential of tumor-agnostic biomarkers to guide effective therapies, randomized evidence supporting the clinical superiority of precision oncology approaches compared to standard therapies remains limited. The ROME trial was a multicenter, randomized, open-label phase 2 study comparing tailored treatment (TT) to standard of care (SoC) in patients with advanced solid tumors progressing after one or two lines of therapy. Comprehensive genomic profiling on tissue and blood was performed to identify actionable alterations. Overall response rate (ORR) was the primary endpoint, and progression-free survival (PFS), overall survival (OS), time to treatment failure (TTF), time to next treatment (TTNT) and safety were the secondary endpoints. Between November 2020 and August 2023, 1,794 patients were screened, 897 were evaluated by the molecular tumor board (MTB) and 400 were randomized to TT or SoC. TT achieved a significantly higher ORR (17.5% versus 10%; $P = 0.0294$) and improved median PFS (3.5 months versus 2.8 months; hazard ratio = 0.66 (0.53–0.82), $P = 0.0002$). TT also showed superior 12-month PFS rates (22.0% versus 8.3%). Median OS was similar, with a 52% crossover rate. Grade 3/4 adverse events were also similar (40% TT versus 52% SoC). These results highlight the potential of TT to improve outcomes for patients with diverse actionable genomic alterations. These results also provide relevant evidence supporting a tumor-agnostic precision oncology strategy and highlight the potential of TTs, guided by genomic profiling and MTB recommendations, to significantly improve outcomes for patients with diverse actionable genomic alterations. ClinicalTrials.gov identifier: [NCT04591431](https://clinicaltrials.gov/ct2/show/study/NCT04591431).

Precision oncology aims to enhance clinical outcomes by tailoring cancer treatments to individual patients, focusing on specific vulnerabilities identified through the analysis of cancer characteristics and the patient's clinical history¹. However, implementing these strategies presents major challenges, such as detecting actionable biomarkers, interpreting their clinical significance within the context of an individual patient's history and systematically utilizing available tailored treatments (TTs). This complexity is further compounded by the variability in the frequency of actionable molecular alterations across different tumor types, making histology a less reliable factor for determining

treatment options². Additionally, standard therapeutic options vary considerably across different tumor types. As a result, the clinical significance of an actionable genetic alteration can differ substantially across various cancer types³.

Among the different approaches for detailed molecular characterization of tumor samples, genomic tests are the most widely used in clinical trials to identify patients with actionable alterations.

Recent advancements have uncovered that treatments targeting some tumor-agnostic biomarkers induce considerable clinical benefits regardless of histology. Notable examples include pembrolizumab⁴

✉ e-mail: simone.scagnoli@uniroma1.it

and dostarlimab⁵ for tumors exhibiting high microsatellite instability (MSI-H); pembrolizumab for high tumor mutational burden (hTMB)⁶; larotrectinib⁷, entrectinib⁸ and repotrectinib⁹ for tumors harboring *NTRK* gene fusions; dabrafenib and trametinib for *BRAF* V600 mutation-positive tumors¹⁰; seliprecatinib for *RET* fusions¹¹; and trastuzumab deruxtecan for HER2-overexpressing (3+) solid tumors¹².

However, definitive evidence demonstrating the superiority of these approaches over standard therapies is lacking, with inconsistent outcomes reported across randomized agnostic precision oncology clinical trials. For instance, although the SHIVA trial did not show progression-free survival (PFS) benefits for off-label TT compared to standard treatments¹³, the MOSCATO-01 trial achieved its endpoint, underscoring the value of broad genomic testing and targeted drug treatments¹⁴.

Large, prospective, non-randomized trials are being conducted to identify molecularly defined subpopulations of patients deriving benefit from TTs. The results of multiple cohorts of the Targeted Agent and Profiling Utilization Registry (TAPUR) study were published^{15–18}. Similarly, the MyPathway and the NCI-MATCH trials provided valuable insights into the actionability of molecular findings in multiple tumor types^{19–21}.

The European Society for Medical Oncology (ESMO) sought to address these challenges by developing tools such as ETAC-S, which contains minimum requirements for identifying TTs with tumor-agnostic potential based on mechanistic evidence paired with prospective clinical insights²². Another widely implemented framework is the ESMO Scale for Clinical Actionability of molecular Targets (ESCAT)²³, which ranks genomic aberrations to guide therapeutic decisions. The SAFIRO2-BREAST trial demonstrated the utility of ESCAT, indicating that only patients harboring genomic alterations ranked as tier I and tier II benefit from maintenance TT, whereas those harboring alterations with lower actionability levels do not²⁴.

Moreover, molecular tumor boards (MTBs) provide essential multidisciplinary expertise for implementing precision medicine²⁵. However, the impact of MTBs on patient outcomes requires further validation, as current evidence is limited and varies based on factors such as drug availability and tumor types treated^{26–29}. Despite these advancements, important hurdles remain in effectively implementing precision oncology in Italy³⁰. Although regulatory approvals for targeted therapies have increased recently, many MTB-recommended treatments continue to face narrow or delayed approvals, limiting patient access to TTs even when actionable alterations are detected. Consequently, referral to clinical trials often becomes a viable option for patients to access TTs. Challenges such as limited enrollment availability and strict inclusion criteria further hinder the efficiency of this approach. Therefore, collaborative policies are essential to overcome these issues and facilitate the implementation of precision oncology in clinical practice²⁵.

The nationwide ROME trial was designed to provide comprehensive genomic profiling and TT for patients with solid tumors across multiple medical oncology units in Italy. This trial aims to evaluate the outcomes of a TT established after discussions in the MTB compared to the standard of care (SoC). Additionally, this approach seeks to generate high-quality data to identify potentially effective treatments that deserve further clinical validation in specific biomarker-defined subsets of patients.

Results

Patient enrollment and baseline characteristics

From 13 October 2020 to 19 July 2023, 1,794 patients with Eastern Cooperative Oncology Group Performance Status (ECOG PS) of 0 or 1 and metastatic solid tumors, who had received up to two lines of therapy, were accrued across 40 oncological centers in Italy. These patients were screened for genomic alteration on tissue and peripheral blood with FoundationOne CDx and FoundationOne Liquid CDx centralized

next-generation sequencing (NGS). A total of 897 patients with potentially actionable molecular alterations were identified and discussed during 127 weekly MTB meetings. Patients with mutations deemed actionable by at least one of the drugs available in the trial were subsequently randomized in a 1:1 ratio to receive TT versus SoC. In total, 497 patients were excluded from randomization based on MTB directives outlined in Methods.

Actionable genomic alterations and MTB-selected treatment options for the intention-to-treat (ITT) population are reported in Extended Data Fig. 1.

Following the MTB recommendations, 221 (55%) patients were referred to a targeted therapy, 153 (38%) to immunotherapy and 26 (7%) to a combination of both (Extended Data Fig. 2). Thus, 400 patients were ultimately randomized into the TT and SoC arms. Patients were allowed to crossover at the locally confirmed progression of the disease. The CONSORT diagram is presented in Fig. 1.

Patient characteristics and lines of prior therapy are detailed in Table 1 and Supplementary Table 1, respectively. The median age, baseline ECOG PS and number of previous lines of treatment were similar between arms. The TT arm had a higher proportion of women. The primary tumor origins were well balanced, with only glioblastomas and biliary tract tumors showing a slight overrepresentation in the SoC and TT arms, respectively, with no statistical differences observed. The TT arm also had a higher percentage of patients who received prior targeted therapy or hormonal therapy or a combination of chemotherapy and targeted therapy. In contrast, the SoC arm had more patients with prior immunotherapy.

Efficacy and survival outcomes

The overall response rate (ORR) in the ITT population of 400 patients demonstrated a statistically significant improvement in the TT arm compared to the SoC arm. Among the 200 patients receiving TT, the ORR was 17.5% (95% confidence interval (CI): 12.5–23.5), which included 3% (six patients) achieving a complete response and 14.5% (29 patients) achieving a partial response. In contrast, the SoC arm achieved an ORR of 10% (95% CI: 6.2–15.0), with no complete responses and a 10% partial response rate (20 patients). Stable disease was observed in 19.5% (39 patients) in the TT arm and in 17.5% (35 patients) in the SoC arm, and progressive disease was reported in 63.0% and 72.5% of patients in the TT and SoC arms, respectively. The primary endpoint analysis conducted using the stratified analysis by Cochran–Mantel–Haenszel (CMH) test demonstrated a significant difference in ORR between groups ($P = 0.0285$; Extended Data Table 1). ORR varied across pre-specified strata: in the breast cancer subgroup (Stratum A), ORR was 20.0% for TT versus 35.0% for SoC; in the non-colorectal gastrointestinal cohort (Stratum B), rates were 14.8% versus 9.1%; in non-small cell lung cancer (NSCLC) (Stratum C), 12.5% versus 6.7%; and in ‘Other malignancies’ (Stratum D), 19.1% versus 6.4%. The unique discrepancy with general results was observed in the breast cancer subgroup, probably as a consequence of the small breast cancer cohort and its inherent heterogeneity. The P value from the chi-square test ($P = 0.0294$; Extended Data Table 2) is nearly identical to the stratified CMH result, supporting the robustness of the main conclusion of the study. Furthermore, the Breslow–Day test for homogeneity of odds ratios across strata was not significant ($P = 0.1231$), failing to provide evidence of heterogeneity in the treatment effect across cancer types. PFS was significantly improved in the TT arm (median 3.5 months, 95% CI: 3.0–4.8) compared to the SoC arm (2.8 months, 95% CI: 2.5–3.2), with a hazard ratio (HR) of 0.66 (95% CI: 0.53–0.82, $P = 0.0002$; Fig. 2a). This benefit extended to key timepoints, showing substantial higher 9-month and 12-month PFS rates in the TT arm: 27.8% (95% CI: 21.4–34.2) at 9 months and 22.0% (95% CI: 16.0–28.0) at 12 months compared to 13.4% (95% CI: 8.4–18.5) and 8.3% (95% CI: 4.2–12.5), respectively, in the SoC arm. No significant difference in median overall survival (mOS) was observed between the TT and SoC arms (mOS: 9.1 months (TT) and 7.9 months

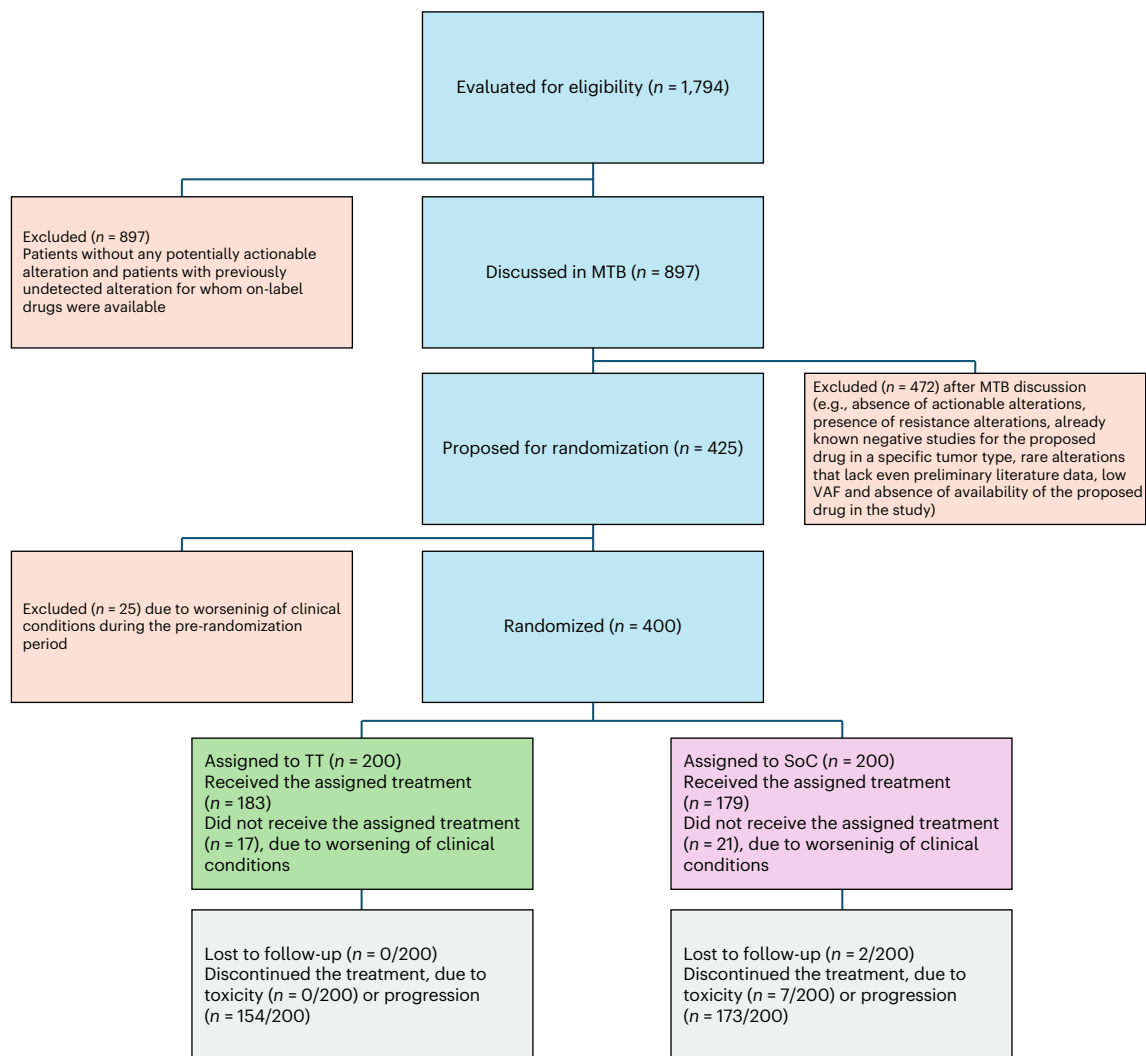


Fig. 1 | CONSORT diagram. CONSORT diagram for the ROME trial.

(SoC); HR = 0.92, 95% CI: 0.72–1.19, $P = 0.5322$; Fig. 2b). A high crossover rate (59%) from SoC to TT occurred after disease progression. Among non-crossover patients in the SoC arm ($n = 74$), key reasons included rapid clinical deterioration ($n = 33$) and death before crossover eligibility ($n = 25$). Full details are reported in Supplementary Tables 2 and 3. Approximately half of the enrolled patients in both arms had already received two prior lines of systemic therapy, which may have influenced both treatment responsiveness and overall clinical outcome. Analyses of the key secondary endpoints—time to treatment failure (TTF) and time to next treatment (TTNT)—showed superior outcomes in the TT arm compared to the SoC arm. Median TTF was 3.5 months (95% CI: 3.1–4.9) in the TT arm versus 2.8 months (95% CI: 2.5–3.1) in the SoC arm (HR = 0.64, 95% CI: 0.51–0.79, $P < 0.0001$). At 12 months, TTF was 22.0% (95% CI: 16.0–28.0) in the TT arm and 8.6% (95% CI: 4.4–12.7) in the SoC arm (Fig. 2c). Patients receiving TT had a significantly longer TTNT than those in the SoC arm (median TTNT: 5.0 months (95% CI: 3.9–6.3) versus 3.5 months (95% CI: 3.0–3.8); HR = 0.59, 95% CI: 0.47–0.75). The TT arm also demonstrated a significantly higher proportion of patients remaining on treatment at 12 months (24.6%, 95% CI: 18.1–31.1) than the SoC arm (8.7%, 95% CI: 4.4–13.0) (Fig. 2d).

Safety

The overall incidence of grade ≥ 3 adverse events (AEs) was similar in the TT and SoC arms (40.0% (80/200) and 52.5% (105/200), respectively). However, the specific AEs differed between the arms. In the TT

arm, diarrhea was the most common AE (11 patients, 5.5%), whereas neutropenia was the most common AE in the SoC arm (16 patients, 8.0%) (Fig. 3 and Extended Data Fig. 3). Figure 3 presents a tornado plot illustrating these AEs, categorized by System Organ Class categories of the Common Terminology Criteria for Adverse Events (CTCAE) version 5. Extended data provide a comprehensive list of grade ≥ 3 AEs (Extended Data Fig. 3a) and all grade 5 AEs (Extended Data Fig. 3b).

Exploratory subgroup analyses

Further exploratory analyses in specific subgroups provided additional insights. To assess the efficacy of TT against SoC, we analyzed subgroups defined by clinically relevant molecular alterations: MSI-H, hTMB, *BRAF* mutations and *ERBB2* (*HER2*) alterations (including amplifications and mutations). Within the ITT population, these alterations were detected with the following frequencies: hTMB/microsatellite stable (MSS) (≥ 10 mutations per megabase, 33.7%), MSI-H (4.5%), *BRAF* mutations (4.3%) and *ERBB2* alterations (14.3%). Treatment allocation was as follows: patients with hTMB, irrespective of microsatellite status, in the TT arm received immunotherapy (ipilimumab plus nivolumab or nivolumab monotherapy); patients with *BRAF* mutations received vemurafenib plus cobimetinib; and patients with *ERBB2* alterations received one of several anti-HER2 regimens (pertuzumab plus trastuzumab, TDM1, trastuzumab plus lapatinib, pertuzumab plus TDM1, TDM1 plus atezolizumab or trastuzumab plus everolimus). The results of these exploratory subgroup analyses of the ORR are presented in

Table 1 | Characteristics of the ITT study population

Characteristics	Overall n=400 (%)	SoC n=200 (%)	TT n=200 (%)
Age (years)			
Median (range)	61 (22–85)	60 (34–84)	62 (22–85)
Gender			
Male	192 (48.0)	100 (50.0)	92 (46.0)
Female	208 (52.0)	100 (50.0)	108 (54.0)
Ethnicity			
Caucasian	392 (98.1)	196 (98.0)	196 (98.0)
Asian	2 (0.5)	0 (0.0)	2 (1.0)
Hispanic	3 (0.8)	2 (1.0)	1 (0.5)
African American	1 (0.2)	1 (0.5)	0 (0.0)
Other	1 (0.2)	0 (0.0)	1 (0.5)
NA	1 (0.2)	1 (0.5)	0 (0.0)
Primary tumor			
Colorectal	65 (16.2)	31 (15.5)	34 (17.0)
Breast	40 (10.0)	20 (10.0)	20 (10.0)
Gastric	36 (9.0)	19 (9.5)	17 (8.5)
Glioblastoma	36 (9.0)	21 (10.5)	15 (7.5)
Biliary tract/gallbladder	35 (8.8)	10 (5.0)	25 (12.5)
NSCLC	35 (8.8)	18 (9.0)	17 (8.5)
Ovarian	18 (4.5)	8 (4.0)	10 (5.0)
Pancreatic	18 (4.5)	10 (5.0)	8 (4.0)
Melanoma	13 (3.2)	8 (4.0)	5 (2.5)
Anus	11 (2.8)	5 (2.5)	6 (3.0)
Others ^a	93 (23.2)	50 (25.0)	43 (21.5)
ECOG PS			
0	237 (59.3)	116 (58.0)	121 (60.5)
1	162 (40.4)	83 (41.5)	79 (39.5)
2	1 (0.3)	1 (0.5)	0 (0.0)
Previous treatment lines			
One	204 (51.0)	99 (50.0)	105 (52.0)
Two	196 (49.0)	101 (50.0)	95 (48.0)

Populations were well balanced after randomization. All variables were tested, and no significant difference was detected between treatment arms. ^aThe detailed description of this group is provided in Supplementary Table 4.

Extended Data Table 3; the swimmer plots for duration of response are reported in Extended Data Figs. 4 and 5; and PFS and OS are reported in Fig. 4.

Of the 135 patients with hTMB and MSS status (65 in the SoC arm and 70 in the TT arm), all patients in the TT arm received immunotherapy (ipilimumab plus nivolumab or, in a few cases, nivolumab monotherapy). The ORR was higher in the TT arm (25.7%, 95% CI: 16–37.6) than in the SoC arm (15.4%, 95% CI: 7.6–26.5) (Extended Data Table 3). PFS was significantly longer in the TT arm (3.2 months, 95% CI: 2.5–5.6) compared to the SoC arm (2.8 months, 95% CI: 2.2–4.0) (HR = 0.64, 95% CI: 0.43–0.94; Fig. 4a). At 12 months, the PFS rate was 32.6% (95% CI: 21.3–43.9) in the TT arm versus 8.1% (95% CI: 0.9–15.3) in the SoC arm, representing a 25% absolute improvement in patients alive and progression free at 1 year. OS was 10.6 months (95% CI: 4.2–16.8) in the TT arm and 7.4 months (95% CI: 4.9–14.1) in the SoC arm (HR = 0.96, 95% CI: 0.62–1.49; Fig. 4b).

Among 18 patients with MSI-H, the ORR was 57.1% (95% CI: 18.4–90.1) in the TT arm and 0% in the SoC arm (Extended Data Table 3).

PFS had not been reached in the TT arm at the time of analysis, whereas it was 2.8 months (95% CI: 1.4–3.7) in the SoC arm. Furthermore, at 1 year, 57.1% (95% CI: 20.5–93.8) of patients in the TT arm remained on treatment compared to none in the SoC arm (HR = 0.15, 95% CI: 0.03–0.73; Fig. 4c). The mOS was not reached in the TT arm. However, at 12 months, the OS rate was 71.4% in the TT arm compared to 33.3% and an mOS of 3.7 months in the SoC arm (HR = 0.38; Fig. 4d).

Of the 14 randomized patients with *BRAF* alterations, five received SoC and nine received TT, consisting of a *BRAF*/MEK inhibitor combination. A notable difference in PFS was observed: PFS was not reached in the TT arm, whereas it was 2.0 months (95% CI: 1.6–not evaluable) in the SoC arm (HR = 0.07, 95% CI: 0.01–0.67; Fig. 4e). The ORR was 22.2% (95% CI: 2.8–60.0) in the TT arm compared to 0% in the SoC arm (Extended Data Table 3). Despite the small sample size, the 12-month PFS rate was substantially higher in the TT arm (64.8%, 95% CI: 32.4–97.2) compared to the SoC arm (0%). The TT arm also demonstrated superior OS compared to the SoC arm; mOS was not reached in the TT arm, whereas it was 8.1 months (95% CI: 1.9–not evaluable) in the SoC arm (HR = 0.26, 95% CI: 0.04–1.57; Fig. 4f).

In the subgroup of 58 patients with *HER2* alterations, various anti-*HER2* agents were administered. One patient in this group had an *ERBB3* mutation, was also *HER2* positive and received an indication for anti-*HER2* treatment. The ORR was 13.8% (95% CI: 3.9–31.7) in the TT arm versus 6.9% (95% CI: 0.9–22.8) in the SoC arm (Extended Data Table 3). PFS was significantly longer in the TT arm (4.6 months, 95% CI: 3.0–5.7) compared to the SoC arm (2.6 months, 95% CI: 1.7–3.0) (HR = 0.4, 95% CI: 0.24–0.74; Fig. 4g). Furthermore, at 12 months, 14.4% (95% CI: 1.40–27.50) of patients in the TT arm remained progression free compared to no patients in the SoC arm. Patients with *HER2*-altered tumors receiving TT showed a trend toward higher OS compared to the SoC arm, with an mOS of 10.2 months (95% CI: 6.5–not evaluable) versus 5.6 months (95% CI: 2.8–13.7) (HR = 0.61, 95% CI: 0.32–1.16; Fig. 4h).

Discussion

The ROME trial provides evidence that TT, guided by comprehensive genomic profiling and expert MTB decision-making, significantly improves outcomes in pretreated patients with metastatic disease. The study demonstrated statistically significant improvements in the ORR and prolonged PFS compared to SoC, underscoring the clinical value of molecularly driven precision oncology. The improvement in ORR, the primary endpoint of the study, observed in the TT arm compared to the SoC arm within the ITT population (17.5% versus 10%), is clinically relevant and methodologically robust, although modest in magnitude. Notably, the TT arm achieved complete responses in a patient population with advanced metastatic disease treated in later lines, whereas no complete responses were observed in the SoC arm, underlining the potential efficacy of TTs in achieving deeper and potentially more durable clinical responses. The improvement in ORR was consistent despite variations across predefined cancer type strata. Subgroup analyses revealed pronounced benefit in the ‘Other malignancies’ stratum (19.1% versus 6.4%), positive trends in non-colorectal gastrointestinal (14.8% versus 9.1%) and NSCLC (12.5% versus 6.7%) subgroups and a lower ORR in the TT arm for breast cancer (20.0% versus 35.0%). However, the analyses of the differences in the odds ratios of response across subgroups have merely exploratory importance due to the limited size of most subgroups and, more importantly, because the primary aim of the trial was to assess the feasibility and potential global impact of a tailored therapeutic strategy based on comprehensive genomic profiling. The observed numerical imbalance across the four predefined cohorts resulted from a combination of protocol-driven and real-world factors. Although the initial design anticipated competitive enrollment with balanced strata, the trial’s biomarker-driven framework inherently favored variability in recruitment. A key factor contributing to this disparity was the differential prevalence of actionable genomic alterations across tumor types. Stratum D, which included diverse

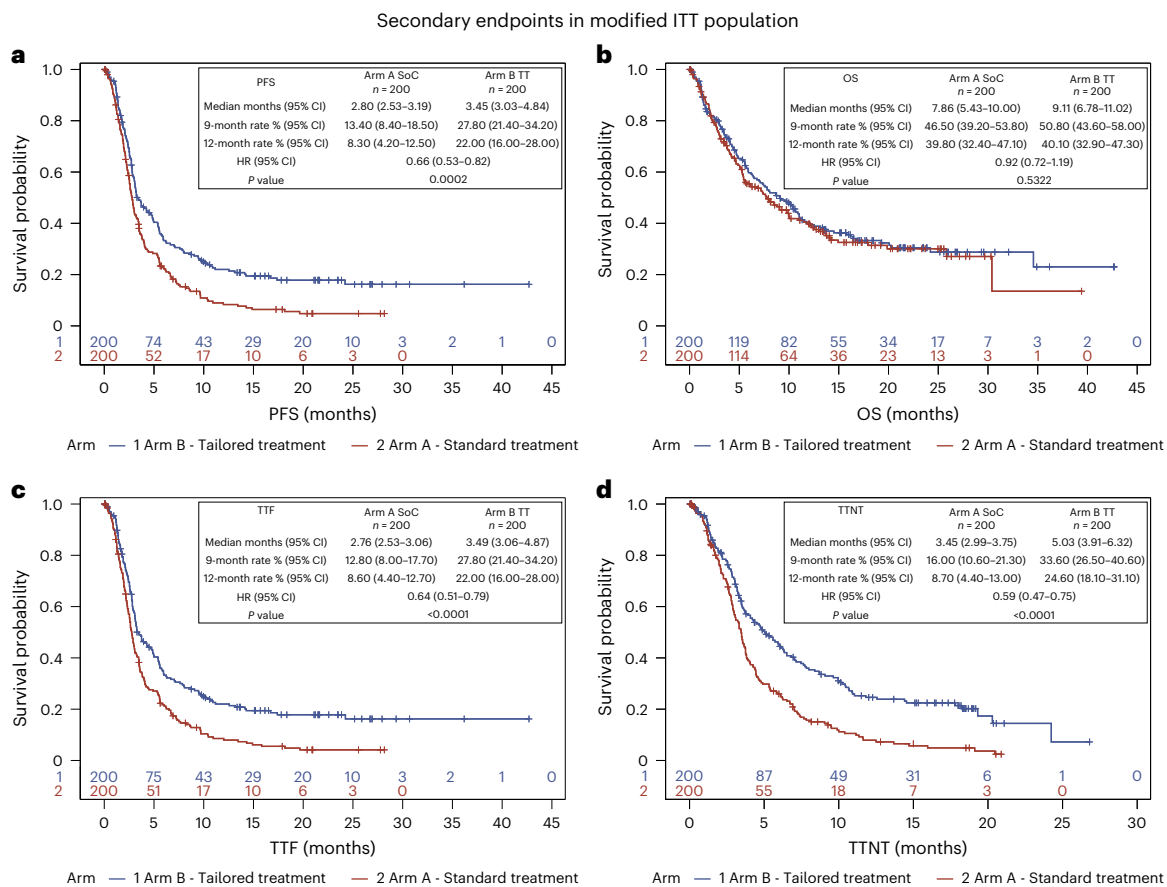


Fig. 2 | Secondary endpoints in the ITT population. Secondary survival endpoints in the ITT population. **a**, PFS ($P = 0.002$). **b**, OS ($P = 0.5322$). **c**, TTF ($P < 0.0001$). **d**, TTNT ($P < 0.0001$). Statistical test used to calculate P values: log-rank test.

malignancies, exhibited higher rates of potentially actionable targets compared to other strata. Molecular profiling success rates further influenced recruitment dynamics. Tumors with lower shed DNA or suboptimal sample quality, common in certain histologies, reduced liquid biopsy utility, whereas tissue availability varied by cancer type. Additionally, site-specific expertise and patient populations probably played an important role, with centers specializing in rare cancers contributing more to Stratum D, whereas those focused on common tumors faced stricter exclusion thresholds. Temporal factors, including profiling turnaround times and drug availability after protocol amendments, also contributed to uneven accrual. Despite these imbalances, the trial's statistical integrity remained robust, as previously discussed. The observed disparities reflect real-world heterogeneity in the distribution of actionable targets rather than methodological shortcomings. The ROME trial's agnostic design, prioritizing molecular over histologic stratification, inherently accommodates such variability, validating its approach in capturing the complexity of precision oncology (Fig. 5). The recruitment pattern underscores a critical challenge in biomarker-driven trials: molecular prevalence and clinical exclusion criteria inevitably influence enrollment dynamics. The trial's outcomes, however, demonstrate that robust statistical methods can mitigate imbalances, ensuring meaningful conclusions even in heterogeneous populations. These insights support the feasibility of tumor-agnostic frameworks while emphasizing the need for adaptive designs in future precision oncology studies. A major methodological departure from prior precision oncology trials is the timing of molecular profiling. In contrast to most studies that also use archival tissue samples from primary tumors, the ROME trial specifically employed metastatic tissue collected upon disease progression after the last therapy (with few exceptions; see supplementary materials). This strategy provided

a more precise evaluation of the tumor's mutational landscape at study entry. The addition of liquid biopsy analysis, comprehensive MTB assessment and early randomization (within the first two lines of therapy) further enhanced the accuracy of TT recommendations while mitigating the confounding effects of clonal heterogeneity frequently observed in later-stage disease.

A key finding from this study is the significant long-term benefit of PFS associated with TT. At the 12-month mark, patients receiving TT demonstrated improved disease control, indicating that it can extend PFS beyond 1 year for certain patients. This benefit, observed even in pre-treated metastatic cases, highlights the clinical importance of personalized treatment strategies based on individual molecular alterations.

The observed significant improvements in ORR and PFS within the TT arm were not accompanied by a similar improvement in OS. The substantial crossover rate (58.8%) from the SoC arm to the TT arm after disease progression, although ethically appropriate, likely contributed to this lack of OS benefit. The high rate of non-crossover (41.2%) underscores the challenges of implementing precision therapies in late-line settings, where declining performance status and emergent complications often preclude treatment changes. This reality suggests that TTs may require earlier deployment to maximize clinical impact. This highlights the need for future studies to incorporate stratified crossover analyses to better understand the long-term effects of TTs on survival outcomes.

Subgroup analyses focused on specific molecular alterations (hTMB, *BRAF* mutations and *ERBB2* alterations) revealed considerable benefits. The hTMB subgroup demonstrated a particularly strong PFS benefit with immunotherapy (nivolumab with or without ipilimumab), showing efficacy across both MSI-H and MSS tumor types. This finding supports hTMB as a predictive biomarker for immunotherapy

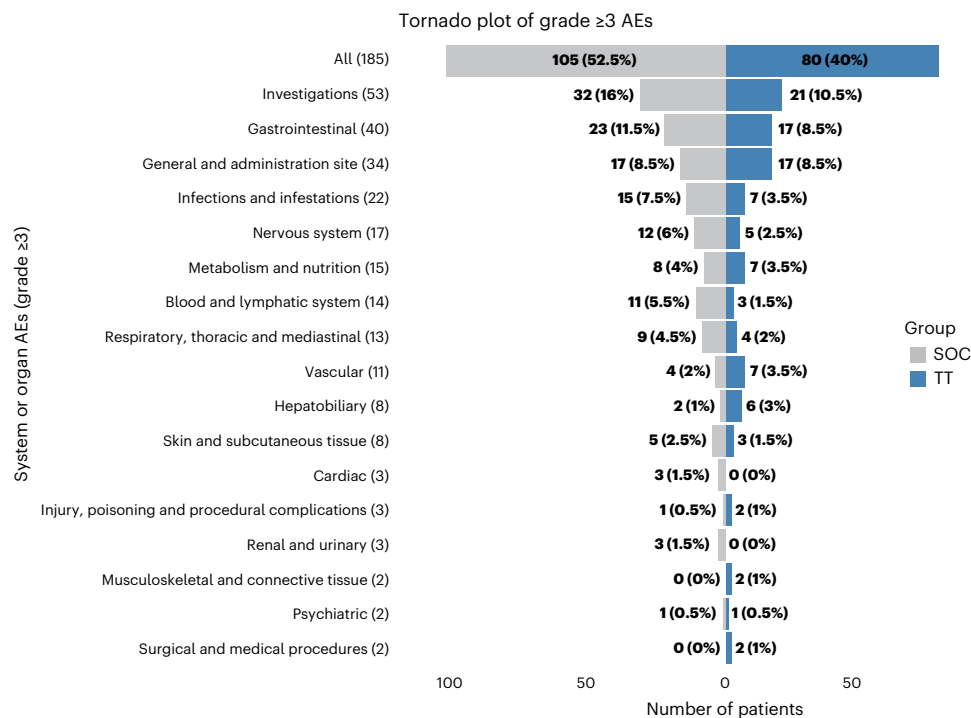


Fig. 3 | Tornado plot with the System Organ Class groups of grade ≥ 3 AEs according to CTCAE version 5. This figure shows the system or organ grade ≥ 3 AEs that occurred during the study for patients receiving SoC or TT.

response, even within a biomarker-agnostic approach, particularly when using the TMB calculation algorithm employed by the Foundation One platform. Despite this promising conclusion, literature reports on TMB have been inconsistent, likely due to several factors. Key contributing elements include methodological variations, site-specific influences and a lack of standardization across the field. The complexity is further compounded by the use of different platforms and proprietary algorithms as well as variability in panel size, gene coverage, bioinformatics pipelines, sequencing depths and tumor purity. Additionally, cancer-type-specific variations play an important role in the assessment of TMB. Despite these challenges, the potential of hTMB as a predictive tool for immunotherapy response remains promising, highlighting the need for continued exploration and standardization efforts. Similarly, TTs showed efficacy against tumors harboring *BRAF* and *HER2* alterations. Although the smaller sample sizes in some subgroups may have limited the statistical power of certain observations, the overall findings emphasize the potential of precision oncology.

The similar incidence of grade 3/4 AEs in the TT and SoC arms, albeit with different AE profiles (diarrhea and fatigue in TT versus neutropenia in SoC), underscores the importance of proactive, personalized management of toxicities to optimize the therapeutic benefit–risk ratio.

The pivotal role of the MTB in shaping clinical decision-making was evident throughout the trial. Composed of a multidisciplinary team of specialists, the MTB enabled a mutation-agnostic and tumor-agnostic approach, tailoring treatment plans based on comprehensive molecular profiles rather than histology alone, avoiding the passive acceptance of the mutation/drug axiom. This approach, combined with the clinical data provided by referring clinicians, facilitated a patient-specific therapeutic strategy. In scenarios where multiple targeted treatment options were available for a single molecular pathway (such as PI3K/AKT or ERBB2 alterations), the MTB prioritized agents based on each patient's unique molecular profile and clinical characteristics. Factors driving these decisions included the specific type of alteration (for example, *ERBB2* mutation versus amplification), published evidence on drug efficacy in specific contexts, prior

therapeutic exposures and drug safety profiles. Notably, several agents (alpelisib, tepotinib, talazoparib, seliparitinib and pralsetinib) were introduced via protocol amendment in 2024. These updates, documented in Extended Data Table 4, preserved trial integrity through predefined statistical analysis plan (SAP) stratification while enhancing real-world relevance.

The MTB's ability to address each patient's biological complexity and clinical heterogeneity optimized therapy selection, contributing substantially to the positive outcomes of this trial. In case of detection of multiple actionable alterations, the MTB evaluated the potential clinical benefit of each therapeutic option, considering evidence from published literature, drug mechanism of action, patient-specific clinical factors, prior therapeutic responses and toxicity risks. The presence of multiple actionable biomarkers considerably complicates therapeutic decision-making due to pathway interactions, conflicting clinical evidence and diverse drug toxicities. Managing this complexity often exceeds individual clinician capacity and simple biomarker–drug algorithms. In the era of precision oncology, the MTB model is proving essential for interpreting complex genomic data and guiding personalized treatment plans that extend beyond traditional histopathological and direct target–drug association models, as seen in the prescription of TTs based on demonstrated efficacy in controlled clinical studies. The ROME trial provides, to our knowledge, the strongest evidence supporting MTB implementation in clinical practice, thanks to its agnostic and randomized design. Previous evidence suggested a potential improvement in survival outcomes when MTBs were implemented, as published by Miller et al.³¹ in a prospective, non-randomized phase 2 study. MTB utility is also being assessed in other prospective clinical trials, such as the Danish phase 2, non-randomized ProTarget study, implementing a Drug Rediscovery Protocol (DRUP)-like design³². Another example is the TARGET study (NCT04723316), a large nationwide prospective trial based in the United Kingdom designed for this purpose, aiming to enroll 6,000 patients with advanced cancers³³. Our findings underscore the critical need to further develop MTBs and implement advanced decision support tools to effectively integrate and interpret complex multiomic data, ultimately enhancing the precision

Exploratory subgroup analysis

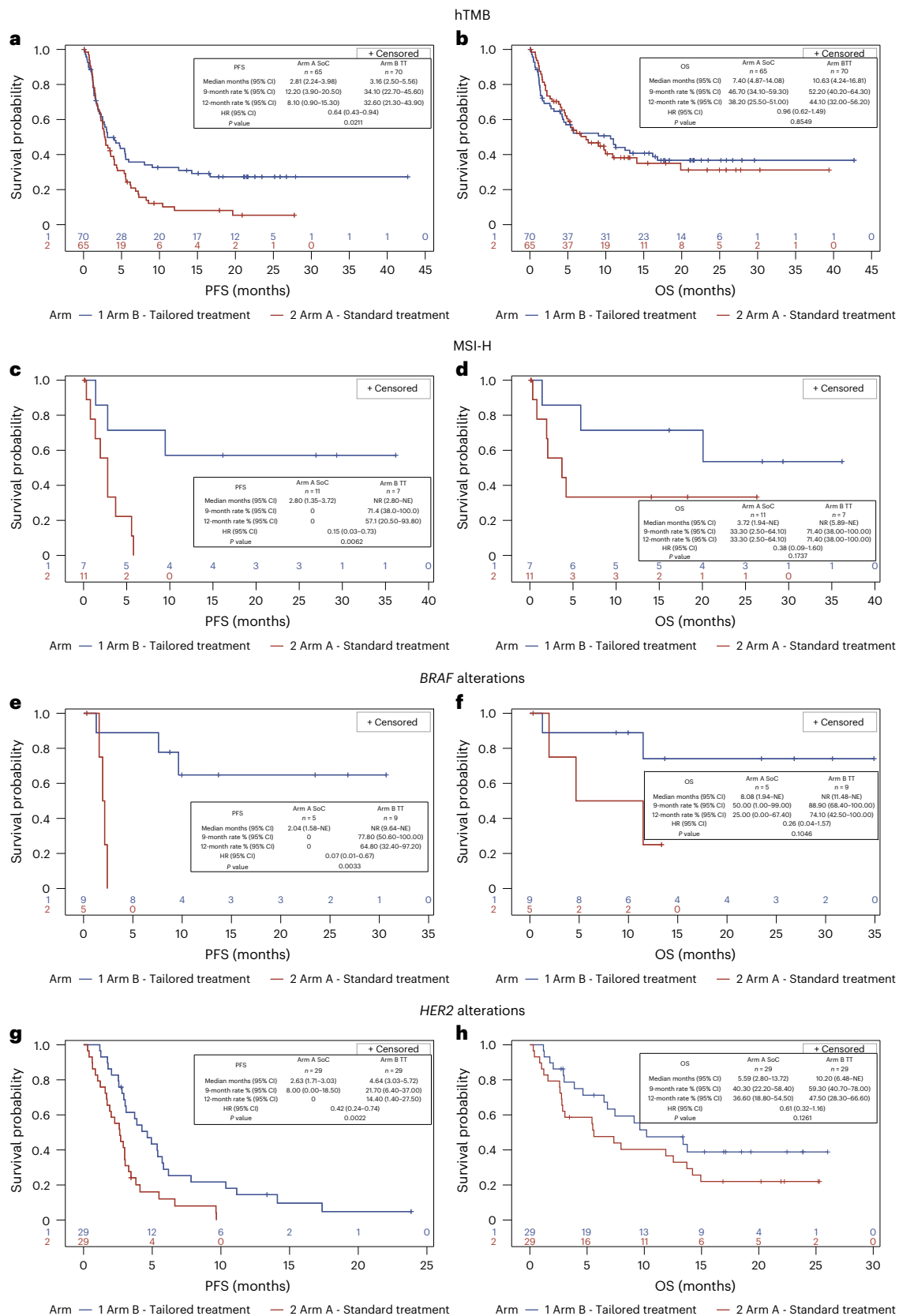


Fig. 4 | PFS and OS in exploratory subgroups. **a**, PFS in hTMB ($P = 0.0211$). **b**, OS in hTMB ($P = 0.8549$). **c**, MSI-H PFS ($P = 0.0062$). **d**, MSI-H OS ($P = 0.1737$). **e**, BRAF alterations PFS ($P = 0.0033$). **f**, BRAF alterations OS ($P = 0.1046$). **g**, HER2 alterations PFS ($P = 0.0022$). **h**, HER2 alterations OS ($P = 0.1261$). Statistical test used to calculate P values: log-rank test. P values derived from exploratory analyses should

be interpreted with caution, as these analyses are hypothesis generating rather than confirmatory. Due to the non-predefined nature of exploratory endpoints and the potential for an inflated α level, the statistical significance observed in these analyses serves primarily to generate hypotheses for future confirmatory studies rather than providing definitive evidence NE, not evaluable; NR, not reached.

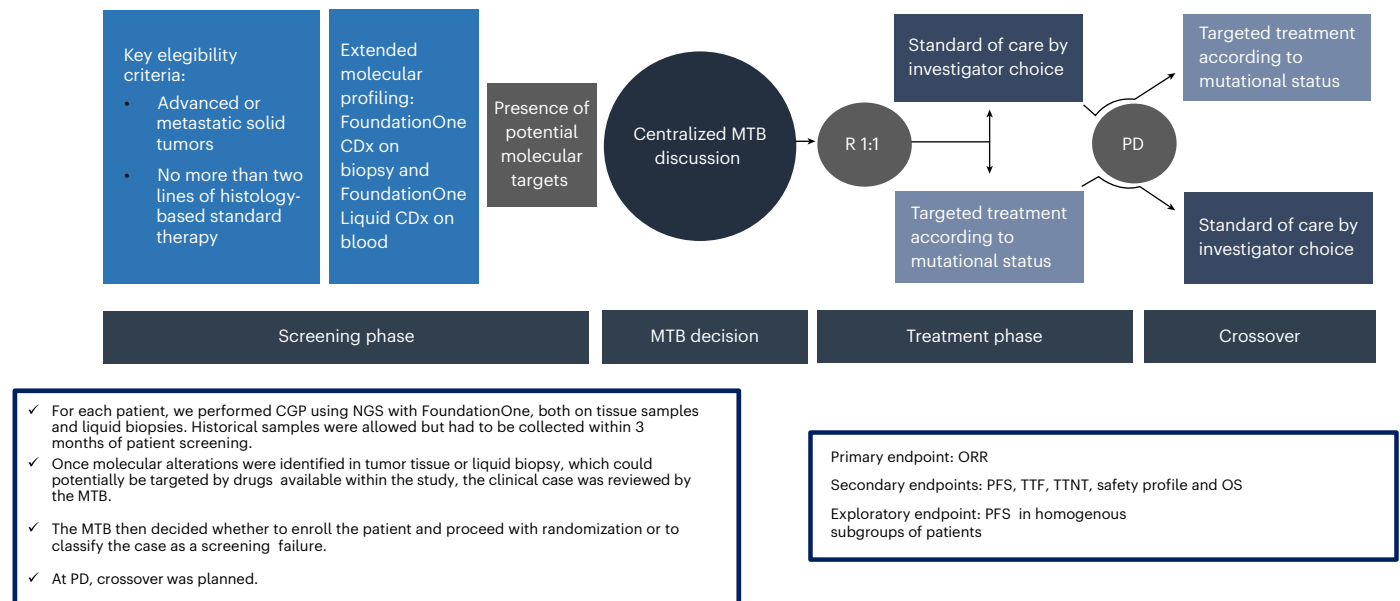


Fig. 5 | Schematic representation of the ROME trial design. The patient flow is shown from the left to right of the figure. The upper boxes represent the two main eligibility criteria, molecular profiling, MTB evaluation, randomization, treatment with TT or SoC and possible crossover at disease progression.

The central blue boxes indicate the study phases. The lower boxes summarize the main study characteristics (left) and the study endpoints (right). CGP, comprehensive genomic profiling; PD, progressive disease; R, randomization.

of clinical recommendations. The management of dual biomarkers (that is, hTMB and *HER2* alteration) posed unique challenges reflecting real-world precision oncology dilemmas, where pathway crosstalk (for example, MAPK activation dampening immunotherapy efficacy) and heterogeneous evidence bases complicate decisions. Our MTB addressed these scenarios by carefully evaluating multiple dimensions, including the biological relevance and actionability of each target (assessed using validated tools), variant allele frequency (VAF) of the detected alterations, quantitative thresholds for biomarkers such as TMB, strength of existing clinical evidence, potential interactions between targeted pathways, patient-specific clinical factors (including prior treatments, clinical status and comorbidities) and safety and toxicity profiles of candidate therapies, thus enabling structured and patient-centered decision-making. AI platforms can enhance MTB decisions in multitarget scenarios by structuring complex genomic/clinical data (for example, prioritizing therapies for *HER2*⁺/hTMB cases) and reducing cognitive overload through evidence-based recommendations. In Italy, the Alliance Against Cancer (ACC) consortium is advancing AI tools under initiatives such as CraNE/EUNetCCC to support MTBs in analyzing mutational profiles, aligning with European Union goals for precision oncology. These systems cross-reference real-world outcomes, biomarker–drug evidence and toxicity profiles, offering clinicians actionable hierarchies while maintaining human oversight. The work of the ACC exemplifies how AI augments, rather than replaces, expert deliberation, improving efficiency and consistency in biomarker-driven care.

The complexities of precision oncology necessitate adaptive platform trials to expedite patient access to effective TTs and keep pace with rapidly advancing genomic knowledge^{34,35}. Platform trials, often referred to as multiarm, multistage studies, are randomized controlled trials (RCTs) that evaluate multiple treatments within different arms of a single trial, all compared to a shared control arm. This design incorporates adaptive features, allowing for the removal of ineffective treatment arms and the inclusion of new arms tailored to specific drug–target interactions. Additionally, patients may be reassigned to different treatment arms based on their response to prior therapies or changes in biomarker profiles. By replacing the traditional sequential trial model, this approach accelerates the process of drug discovery

and development³⁶. Furthermore, it will be beneficial to complement RCTs with observational studies to assess the generalizability and real-world applicability of their results. Future precision oncology will strongly benefit from dynamic national and international clinical trial databases, comprehensively annotated with gene-specific and variant-specific targets, directly linked to advanced decision support platforms to facilitate patient stratification into molecularly driven trials. Although the ROME trial provides robust evidence regarding the efficacy and safety of molecularly guided therapies in a controlled environment, the complexities of real-world practice—including patient heterogeneity, varied access to genomic testing and differences in healthcare infrastructure—may influence the outcomes observed. Observational studies using robust real-world data sources, such as population-based cancer registries, electronic health records and multicenter collaborative networks, combined with advanced AI analysis, would be instrumental in addressing these aspects. Future research priorities include optimizing biomarker-driven patient selection and MTB processes, developing innovative trial designs, investigating specific strategies to overcome resistance mechanisms and exploring combination approaches with immunotherapy and other tailored agents to propel the field of precision oncology further and realize its potential.

Online content

Any methods, additional references, Nature Portfolio reporting summaries, source data, extended data, supplementary information, acknowledgements, peer review information; details of author contributions and competing interests; and statements of data and code availability are available at <https://doi.org/10.1038/s41591-025-03918-x>.

References

1. Prasad, V., Fojo, T. & Brada, M. Precision oncology: origins, optimism, and potential. *Lancet Oncol.* **17**, e81–e86 (2016).
2. Blum, A., Wang, P. & Zenklusen, J. C. SnapShot: TCGA-analyzed tumors. *Cell* **173**, 530 (2018).
3. Schettini, F. et al. Next-generation sequencing-based evaluation of the actionable landscape of genomic alterations in solid tumors: the ‘MOZART’ prospective observational study. *Oncologist* **30**, oyae206 (2025).

4. Marabelle, A. et al. Efficacy of pembrolizumab in patients with noncolorectal high microsatellite instability/mismatch repair-deficient cancer: results from the phase II KEYNOTE-158 study. *J. Clin. Oncol.* **38**, 1–10 (2020).
5. André, T. et al. Antitumor activity and safety of dostarlimab monotherapy in patients with mismatch repair deficient solid tumors: a nonrandomized controlled trial. *JAMA Netw. Open* **6**, E2341165 (2023).
6. Marabelle, A. et al. Association of tumour mutational burden with outcomes in patients with advanced solid tumours treated with pembrolizumab: prospective biomarker analysis of the multicohort, open-label, phase 2 KEYNOTE-158 study. *Lancet Oncol.* **21**, 1353–1365 (2020).
7. Hong, D. S. et al. Larotrectinib in patients with TRK fusion-positive solid tumours: a pooled analysis of three phase 1/2 clinical trials. *Lancet Oncol.* **21**, 531–540 (2020).
8. Drilon, A. et al. Safety and antitumor activity of the multitargeted pan-TRK, ROS1, and ALK inhibitor entrectinib: combined results from two phase I trials (ALKA-372-001 and STARTRK-1). *Cancer Discov.* **7**, 400–409 (2017).
9. Drilon, A. et al. Repotrectinib in ROS1 fusion-positive non-small-cell lung cancer. *N. Engl. J. Med.* **390**, 118–131 (2024).
10. Gouda, M. A. & Subbiah, V. Expanding the benefit: dabrafenib/trametinib as tissue-agnostic therapy for BRAF V600E-positive adult and pediatric solid tumors. *Am. Soc. Clin. Oncol. Educ. Book* **43**, e404770 (2023).
11. Subbiah, V. et al. Tumour-agnostic efficacy and safety of selpercatinib in patients with RET fusion-positive solid tumours other than lung or thyroid tumours (LIBRETTO-001): a phase 1/2, open-label, basket trial. *Lancet Oncol.* **23**, 1261–1273 (2022).
12. Meric-Bernstam, F. et al. Efficacy and safety of trastuzumab deruxtecan in patients with HER2-expressing solid tumors: primary results from the DESTINY-PanTumor02 phase II trial. *J. Clin. Oncol.* **42**, 47–58 (2024).
13. Le Tourneau, C. et al. Molecularly targeted therapy based on tumour molecular profiling versus conventional therapy for advanced cancer (SHIVA): a multicentre, open-label, proof-of-concept, randomised, controlled phase 2 trial. *Lancet Oncol.* **16**, 1324–1334 (2015).
14. Massard, C. et al. High-throughput genomics and clinical outcome in hard-to-treat advanced cancers: results of the MOSCATO 01 trial. *Cancer Discov.* **7**, 586–595 (2017).
15. Mangat, P. K., Garrett-Mayer, E., Perez, J. K. & Schilsky, R. L. The Targeted Agent and Profiling Utilization Registry Study: a pragmatic clinical trial. *Clin. Trials* **20**, 699–707 (2023).
16. Schuetze, S. et al. Palbociclib in patients with soft tissue sarcoma with CDK4 amplifications: results from the Targeted Agent and Profiling Utilization Registry Study. *JCO Precis. Oncol.* **8**, e2400219 (2024).
17. Gupta, R. et al. Pertuzumab plus trastuzumab in patients with colorectal cancer with ERBB2 amplification or ERBB2/3 mutations: results from the TAPUR study. *JCO Precis. Oncol.* **6**, e2200306 (2022).
18. Alva, A. S. et al. Pembrolizumab in patients with metastatic breast cancer with high tumor mutational burden: results from the Targeted Agent and Profiling Utilization Registry (TAPUR) study. *J. Clin. Oncol.* **39**, 2443–2451 (2021).
19. Meric-Bernstam, F. et al. Pertuzumab plus trastuzumab for HER2-amplified metastatic colorectal cancer (MyPathway): an updated report from a multicentre, open-label, phase 2a, multiple basket study. *Lancet Oncol.* **20**, 518–530 (2019).
20. Kalinsky, K. et al. Effect of capivasertib in patients with an AKT1 E17K-mutated tumor: NCI-MATCH subprotocol EAY131-Y nonrandomized trial. *JAMA Oncol.* **7**, 271–278 (2021).
21. Flaherty, K. T. et al. The Molecular Analysis for Therapy Choice (NCI-MATCH) trial: lessons for genomic trial design. *J. Natl Cancer Inst.* **112**, 1021–1029 (2020).
22. Westphalen, C. B. et al. The ESMO Tumour-Agnostic Classifier and Screener (ETAC-S): a tool for assessing tumour-agnostic potential of molecularly guided therapies and for steering drug development. *Ann. Oncol.* **35**, 936–953 (2024).
23. Mateo, J. et al. A framework to rank genomic alterations as targets for cancer precision medicine: the ESMO Scale for Clinical Actionability of molecular Targets (ESCAT). *Ann. Oncol.* **29**, 1895–1902 (2018).
24. Andre, F. et al. Genomics to select treatment for patients with metastatic breast cancer. *Nature* **610**, 343–348 (2022).
25. Marchetti, P. et al. Do more targets allow more cancer treatments, or not? *Eur. J. Cancer* **187**, 99–104 (2023).
26. Repetto, M. et al. Molecular tumour board at European Institute of Oncology: report of the first three year activity of an Italian precision oncology experience. *Eur. J. Cancer* **183**, 79–89 (2023).
27. Vingiani, A. et al. Molecular tumor board as a clinical tool for converting molecular data into real-world patient care. *JCO Precis. Oncol.* **7**, e2300067 (2023).
28. Tommasi, S. et al. The molecular tumor board as a step in cancer patient management: a southern Italian experience. *Front. Med. (Lausanne)* **11**, 1432628 (2024).
29. Giacomini, P. et al. The Molecular Tumor Board of the Regina Elena National Cancer Institute: from accrual to treatment in real-world. *J. Transl. Med.* **21**, 725 (2023).
30. Crimini, E. et al. Challenges and obstacles in applying therapeutical indications formulated in molecular tumor boards. *Cancers (Basel)* **14**, 3193 (2022).
31. Miller, R. W. et al. Molecular tumor board-assisted care in an advanced cancer population: results of a phase II clinical trial. *JCO Precis. Oncol.* **6**, e2100524 (2022).
32. Kringelbach, T. et al. ProTarget: a Danish Nationwide Clinical Trial on Targeted Cancer Treatment based on genomic profiling—a national, phase 2, prospective, multi-drug, non-randomized, open-label basket trial. *BMC Cancer* **23**, 182 (2023).
33. Ortega-Franco, A. et al. TARGET National: a U.K.-wide liquid-based molecular profiling program to enhance recruitment to early-phase trials. *J. Clin. Oncol.* **40**, TPS3163–TPS3163 (2022).
34. Stenzinger, A. et al. Implementation of precision medicine in healthcare—a European perspective. *J. Intern. Med.* **294**, 437–454 (2023).
35. Gnant, M. et al. The future of clinical trials—goals, ideas, and discussion. In *memo – Magazine of European Medical Oncology* **17**, 77–86 (2024).
36. Haj Mohammad, S. F. et al. The evolution of precision oncology: the ongoing impact of the Drug Rediscovery Protocol (DRUP). *Acta Oncol.* **63**, 368–372 (2024).

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Paolo Marchetti¹, Giuseppe Curigliano^{2,3}, Mauro Biffoni⁴, Sara Lonardi⁵, Simone Scagnoli^{6,7}✉, Lorenzo Fornaro⁸, Valentina Guarneri^{5,9}, Ugo De Giorgi¹⁰, Paolo Antonio Ascierto¹¹, Giovanni Blandino¹², Giulia D'Amati¹³, Massimo Aglietta¹³, Chiara Cremolini¹⁴, Pierfranco Conte^{13,15,16}, Edoardo Crimini^{13,16}, Maurizio Ceracchi^{17,18}, Simona Pisegna¹⁹, Sofia Verkhovskaia¹, Roberto Bordonaro²⁰, Sergio Bracarda²¹, Giovanni Butturini²², Lucia Del Mastro^{23,24}, Andrea DeCensi²⁵, Agnese Fabbri²⁶, Elisabetta Fenocchio¹³, Stefania Gori²⁷, Giulio Metro²⁸, Annamaria Pessino²⁹, Daniele Pozzessere³⁰, Fabio Puglisi^{31,32}, Stefano Tamberi³³, Alberto Zambelli^{34,35}, Donatella Marino³⁶, Ettore Capoluongo³⁷, Federico Cappuzzo³⁸, Bruna Cerbelli⁶, Giuseppe Giannini³⁹, Umberto Malapelle⁴⁰, Federica Mazzuca⁴¹, Marianna Nuti⁴², Giancarlo Pruneri⁴³, Maurizio Simmaco^{44,45}, Lidia Strigari⁴⁶, Giuseppe Tonini^{47,48}, Nello Martini⁴⁹ & Andrea Botticelli^{6,7} on behalf of the ROME trial investigators consortia*

¹Department of Oncology, IDI-IRCCS, Rome, Italy. ²European Institute of Oncology, IRCCS, Milan, Italy. ³Department of Oncology and Hemato-Oncology, University of Milan, Milan, Italy. ⁴Department of Oncology and Molecular Medicine, Italian National Institute of Health (ISS), Rome, Italy. ⁵Department of Oncology, Veneto Institute of Oncology IOV-IRCCS, Padua, Italy. ⁶Department of Radiological, Oncological and Pathological Sciences, Sapienza University of Rome, Rome, Italy. ⁷Oncology Unit, AOU Policlinico Umberto I, Rome, Italy. ⁸Department of Translational Research and New Technologies in Medicine and Surgery, University of Pisa, and Division of Medical Oncology, Pisa University Hospital, Pisa, Italy. ⁹Department of Surgery, Oncology, and Gastroenterology, University of Padua, Padua, Italy. ¹⁰IRCCS Istituto Romagnolo per lo Studio dei Tumori (IRST) 'Dino Amadori', Meldola, Italy. ¹¹Istituto Nazionale Tumori IRCCS Fondazione 'G. Pascale', Naples, Italy. ¹²Translational Oncology Research Unit, IRCCS Istituto Nazionale Tumori Regina Elena (IRE), Rome, Italy. ¹³Candiolo Cancer Institute FPO Istituto di Ricovero e Cura a Carattere Scientifico Candiolo, Candiolo, Italy. ¹⁴Department of Translational Research and New Technologies in Medicine and Surgery, University of Pisa, Pisa, Italy. ¹⁵S. Camillo Hospital IRCCS, Lido di Venezia, Italy. ¹⁶Fondazione Periplo, Cremona, Italy. ¹⁷CMV STAT S.r.l., Rome, Italy. ¹⁸STS, Rome, Italy. ¹⁹Department of Experimental Medicine, Sapienza University of Rome, Rome, Italy. ²⁰Medical Oncology Unit, ARNAS Garibaldi, Catania, Italy. ²¹Oncologia Medica Traslationale, Azienda Ospedaliera Santa Maria di Terni, Terni, Italy. ²²Department of Hepatopancreatobiliary (HPB) Surgery, Pederzoli Hospital, Peschiera del Garda, Verona, Italy. ²³IRCCS Ospedale Policlinico San Martino, UO Clinica Oncologia Medica, Genoa, Italy. ²⁴Department of Internal Medicine and Medical Specialties (DIMI), Università di Genova, Genoa, Italy. ²⁵Department of Medicine and Medical Oncology Unit, E.O. Ospedali Galliera, Genoa, Italy. ²⁶Medical Oncology and Breast Unit, Department of Oncology and Hematology, Central Hospital of Belcolle, Viterbo, Italy. ²⁷Department of Oncology, IRCCS Sacro Cuore Don Calabria Hospital, Negrar di Valpolicella, Verona, Italy. ²⁸Medical Oncology, Santa Maria della Misericordia Hospital, University of Perugia, Perugia, Italy. ²⁹Oncologia Medica 1, IRCCS Ospedale Policlinico San Martino, Genoa, Italy. ³⁰Medical Oncology Unit, Nuovo Ospedale di Prato-Santo Stefano, Azienda USL Toscana Centro, Prato, Italy. ³¹Department of Medical Oncology, CRO Aviano, National Cancer Institute, IRCCS, Aviano, Italy. ³²Department of Medicine, University of Udine, Udine, Italy. ³³Oncology Unit, Santa Maria delle Croci Hospital, AUSL Romagna, Ravenna, Italy. ³⁴Oncology Unit, ASST Papa Giovanni XXIII, Bergamo, Italy. ³⁵Department of Medicine and Surgery, Università degli Studi di Milano Bicocca, Milan, Italy. ³⁶Department of Oncology, Ordine Mauriziano Hospital, Turin, Italy. ³⁷Dipartimento Di Eccellenza In Medicina Molecolare E Biotecnologie Mediche, Università Federico II, Naples, Italy. ³⁸Division of Medical Oncology 2, IRCCS Regina Elena National Cancer Institute, Rome, Italy. ³⁹Department of Molecular Medicine, Sapienza University of Rome, laboratory affiliated to Istituto Pasteur Italia, Fondazione Cenci-Bolognetti, Rome, Italy. ⁴⁰Department of Public Health, University Federico II of Naples, Naples, Italy. ⁴¹Department of Clinical and Molecular Medicine, Sapienza University, Oncology Unit, Azienda Ospedaliera Universitaria Sant'Andrea, Rome, Italy. ⁴²Department of Experimental Medicine, 'Sapienza' University of Rome, Rome, Italy. ⁴³Department of Advanced Diagnostics, Fondazione IRCCS Istituto Tumori di Milano, University of Milan, Milan, Italy. ⁴⁴Laboratory of Clinical Biochemistry, Sant'Andrea University Hospital, Rome, Italy. ⁴⁵Department of Neurosciences, Mental Health and Sensory Organs (NESMOS), Sapienza University of Rome, Rome, Italy. ⁴⁶Department of Medical Physics, IRCCS Azienda Ospedaliero-Universitaria di Bologna, Bologna, Italy. ⁴⁷Department of Medicine and Surgery, Università Campus Bio-Medico, Rome, Italy. ⁴⁸Medical Oncology, Fondazione Policlinico Universitario Campus Bio-Medico, Rome, Italy. ⁴⁹Fondazione ReS, Rome, Italy.

*A list of authors and their affiliations appears at the end of the paper. ✉e-mail: simone.scagnoli@uniroma1.it

the ROME trial investigators consortia

Antonella Savarese⁵⁰, Vincenzo Adamo⁵¹, Davide Quaresmini⁵², Pierosandro Tagliaferri⁵³, Paola Damiano⁵⁴, Benedetta Urbini⁵⁵, Carmelo Bengala⁵⁶, Elena Zaffignani⁵⁷, Carmine Pinto⁵⁸, Alicia Tosoni⁵⁹, Lucio Buffoni⁶⁰, Evaristo Maiello⁶¹, Claudia Mucciarini⁶², Antonio Russo⁶³, Rossana Berardi⁶⁴, Giuseppe Lombardi⁵, Alessandra Piancastelli¹⁰, Gianluca Masi⁸, Laura Bonanno^{9,65}, Vito Vanella¹¹ & Francesca Mannozi¹⁰

⁵⁰Medical Oncology 1, IRCCS Regina Elena National Cancer Institute, Rome, Italy. ⁵¹Clinical Trial Center, Oncology Department A.O., Papardo-Messina, Italy. ⁵²Medical Oncology Unit, IRCCS Istituto Tumori 'Giovanni Paolo II', Bari, Italy. ⁵³Department of Experimental and Clinical Medicine, Magna Graecia University, Catanzaro, Italy. ⁵⁴Centro Oncologico San Leopoldo Mandic, Isola Tiberina Gemelli isola, Roma, Italy. ⁵⁵Department of Medical Oncology, University Hospital, Ferrara, Italy. ⁵⁶Division of Medical Oncology 1, St. Chiara University Hospital, Pisa, Italy. ⁵⁷Department of Oncology and Hematology, AUSL Piacenza Guglielmo da Saliceto Hospital, Piacenza, Italy. ⁵⁸Medical Oncology Unit, Comprehensive Cancer Centre, AUSL-IRCCS di Reggio Emilia, Reggio Emilia, Italy. ⁵⁹Nervous System Medical Oncology Department, IRCCS Istituto delle Scienze Neurologiche di Bologna, Bologna, Italy. ⁶⁰Medical Oncology, Humanitas Gradenigo, Torino, Italy. ⁶¹Oncology Unit, Fondazione IRCCS Casa Sollievo della Sofferenza, San Giovanni Rotondo, Italy. ⁶²Oncology Unit, Ramazzini Hospital, Azienda Unità Sanitaria Locale Modena (AUSL), Carpi, Italy. ⁶³Dipartimento di Medicina di Precisione in Area Medica, Chirurgica e Critica (Me.Pre.C.C.), Università degli studi di Palermo AOU 'Paolo Giaccone', Palermo, Italy. ⁶⁴Clinica Oncologica, Università Politecnica delle Marche – AOU delle Marche, Ancona, Italy. ⁶⁵Medical Oncology 2, Istituto Oncologico Veneto IOV IRCCS, Padova, Italy.

Methods

Inclusion and ethics

The study was approved by the institutional ethics committee of the coordinating center (Sapienza no. rif. C.E. 5575; February 2020) and by the ethics committee of each participating center. The investigational sites that approved the study protocol were as follows: AOU Policlinico Umberto I, Rome; European Institute of Oncology (IEO), IRCCS, Milan; Veneto Institute of Oncology IOV-IRCCS, Padua; Division of Medical Oncology, Pisa University Hospital; IRCCS Istituto Romagnolo per lo Studio dei Tumori (IRST) 'Dino Amadori', Meldola; Istituto Nazionale Tumori IRCCS Fondazione 'G. Pascale', Naples; IRCCS Istituto Nazionale Tumori Regina Elena (IRE), Rome; IRCCS Istituto di Candiolo, Candiolo; Medical Oncology Unit, ARNAS Garibaldi Catania; Azienda Ospedaliera Santa Maria di Terni, Terni; Pederzoli Hospital, Peschiera del Garda; IRCCS Ospedale Policlinico San Martino, Genoa; Ospedali Galliera, Genoa; Central Hospital of Belcolle, Viterbo; IRCCS Sacro Cuore Don Calabria Hospital, Negrar di Valpolicella; Santa Maria della Misericordia Hospital, Perugia; Nuovo Ospedale di Prato-Santo Stefano, Azienda USL Toscana Centro, Prato; CRO Aviano, National Cancer Institute, IRCCS, Aviano; Santa Maria delle Croci Hospital, AUSL Romagna, Ravenna; Oncology Unit ASST Papa Giovanni XXIII, Bergamo; Ordine Mauriziano Hospital, Turin; AOU Policlinico S. Andrea, Rome; Azienda Ospedaliera Universitaria Federico II, Napoli; Fondazione IRCCS Istituto Tumori di Milano; Fondazione Policlinico Universitario Campus Bio-Medico, Rome; A.O. Papardo-Messina; IRCCS Istituto Tumori 'Giovanni Paolo II', Bari; Policlinico universitario 'Mater Domini', Catanzaro; Centro Oncologico San Leopoldo Mandic, Isola Tiberina Gemelli isola; Rome; University Hospital, Ferrara; Misericordia Hospital, Grosseto; AUSL Piacenza Guglielmo da Saliceto Hospital, Piacenza; Comprehensive Cancer Centre, AUSL-IRCCS di Reggio Emilia, Reggio Emilia; Humanitas Gradenigo, Torino; Fondazione IRCCS Casa Sollievo della Sofferenza, San Giovanni Rotondo; Ramazzini Hospital, Azienda Unità Sanitaria Locale Modena (AUSL), Carpi; AOUP 'Paolo Giaccone', Palermo; and AOU delle Marche, Ancona.

The competent authority, Agenzia Italiana del Farmaco (AIFA), authorized the trial on 8 July 2020 (AIFA/SC/P/76132). The trial is registered on ClinicalTrials.gov with identifier [NCT04591431](#) and in the European Union Drug Regulating Authorities Clinical Trials Database (EudraCT) with number [2018-002190-21](#). The trial adhered to the principles of the Declaration of Helsinki regarding research involving human subjects. Forty-one centers received ethics approval and participated in the study enrollment. All patients signed the specifically conceived informed consent form (ICF).

Study design, population and inclusion/exclusion criteria

This randomized, prospective, multicenter, proof-of-concept phase 2 clinical trial enrolled patients with inoperable advanced or metastatic solid tumors, regardless of histology. Eligible patients had failed at least one, but no more than two, prior lines of systemic therapy. An ECOG PS of 0 or 1 was required as well as being at least 18 years of age at the time of signing the ICF. All patients had to have measurable or evaluable disease defined per Response Evaluation Criteria in Solid Tumors version 1.1 (RECIST 1.1)³⁷ or immune-related response criteria (irRC)³⁸. If clinically indicated, magnetic resonance imaging (MRI) and positron emission tomography (PET) scans could have been performed. PET Response Criteria in Solid Tumors (PERCIST)³⁹ were applied in cases where a baseline PET scan was available. Adequate renal, liver and bone marrow functions were required at baseline. Patients who were candidates for potentially curative surgery or other locoregional therapies were excluded. Those with well-established actionable molecular targets, not detected in previous molecular profiling, for which approved and marketed targeted therapies were available—such as *EGFR* mutations or *ALK* translocations in lung cancer, *BRAF* mutations in melanoma, *KIT* mutations in gastrointestinal stromal tumors, *ERBB2* amplification in breast

cancer or tumor-agnostic biomarker-driven treatments already available in clinical practice—were also excluded. Furthermore, patients with only bone and/or brain metastases, uncontrolled brain disease (untreated and/or symptomatic) or those whose brain metastases had not been monitored for over 2 months were excluded. Patients with concurrent severe and/or uncontrolled medical conditions that could compromise their participation in the study (for example, uncontrolled diabetes, clinically relevant cardiac disease, uncontrolled hypertension, congestive heart failure, ventricular arrhythmias, active ischemic heart disease, myocardial infarction within the last year, chronic liver or renal disease, active gastrointestinal ulceration or severely impaired lung function) were also excluded (see supplementary materials for all inclusion/exclusion criteria).

For genomic testing, a tissue sample obtained during the screening phase or within 6 months before enrollment was required. The biopsy should be performed during the screening period, after the completion of conventional therapy for recurrent/metastatic cancer. Samples collected within 3 months before the patient's ICF signature were acceptable, as were samples collected within 6 months, with prior acceptance by the MTB. Archived tissue samples were accepted for patients with glioblastomas and high-grade malignant gliomas. Patients with only one available biopsy, either liquid or solid, due to a failure of one method during the screening phase remained eligible for inclusion and discussion in the MTB. Conversely, if both tissue and liquid biopsy characterizations failed, patients were classified as screening failures.

Molecular profiling was performed using FoundationOne CDX and FoundationOne Liquid CDX tests on tissue and liquid biopsies. DNA from formalin-fixed, paraffin-embedded tumor tissue samples was isolated using the DNA extraction method and analyzed with the FoundationOne CDx panel to assess alterations across 324 genes, including substitutions, insertions and deletions (indels), copy number alterations (CNAs) and gene rearrangements. Genomic signatures of MSI and TMB were also evaluated. For FoundationOne Liquid CDx, circulating cell-free DNA (cfDNA) was extracted from plasma collected from anticoagulated peripheral whole blood samples using FoundationOne Liquid CDx cfDNA blood collection tubes. The cfDNA was analyzed for substitutions, indels in 311 genes, rearrangements in four genes and CNAs in three genes. Additionally, the test evaluated tumor fraction and genomic signatures such as blood-based tumor mutational burden (bTMB) and MSI-H status.

Upon identification of actionable alterations in either tumor tissue or liquid biopsy, and after MTB discussion, selected patients were assigned to receive immunotherapy, targeted therapy or both based on MTB recommendations. A crossover between the two treatment arms was planned upon progressive disease. The study design is reported in Fig. 5.

MTB activities

An MTB convened weekly to review each eligible patient. The MTB consisted of medical oncologists, pathologists, geneticists, immunologists, bioinformaticians and other relevant specialists. Before the MTB discussion, the steering committee identified patients with potentially actionable genomic alterations. The MTB then reviewed each case, considering comprehensive molecular profiling data alongside the patient's clinical features, such as performance status, comorbidities and concurrent treatments. The treating clinician presented each case selected by the steering committee, highlighting potential actionable therapeutic targets.

Molecular alterations were evaluated using databases such as ClinVAR, OncoKB and COSMIC and the ESMO ESCAT scale. The board employed VAF thresholds of 1% for tissue biopsies and 2% for liquid biopsies. It systematically reviewed potential intrinsic resistance mechanisms, such as *KRAS* mutations in patients considered for PIK3CA or AKT inhibitors. For patients with multiple actionable alterations

(for example, hTMB alongside *HER2* amplification or *BRAF* mutation), the MTB prioritized therapies using a hierarchical framework prioritizing clinical urgency (for example, immunotherapy for MSI-H tumors regardless of co-alterations), followed by biomarker–drug evidence strength (for example, *HER2*-targeted agents over exploratory TMB-directed therapies when both were present) and, finally, considering toxicity overlap avoidance (for example, excluding PI3K inhibitors if preexisting hyperglycemia).

Patients discussed at the MTB who were not randomized were excluded for various reasons, including the absence of target alterations, the presence of resistance mutations, known negative studies related to a specific alteration, rare mutations without literature data, low VAF, unavailability of drugs for specific alterations, clinical considerations, high levels of uncertainty regarding potentially targetable alterations and the presence of established targetable alterations with available standard treatments. In cases suggesting a germline origin, germline testing was recommended.

MTB recommendations included randomizing patients to receive TT, referring them to a geneticist in the presence of somatic alterations with potential germline significance, modifying SoC only when necessary and suggesting enrollment in clinical trials or early drug access programs, when available in Italy. MTB activities are summarized in Extended Data Fig. 6.

Treatment regimen

Patients who met the inclusion and exclusion criteria and received a recommendation for a specific targeted agent or combination therapy from the MTB were randomized in a 1:1 ratio to receive either SoC or TT, according to their molecular profiles. In the SoC arm, patients were treated with standard chemotherapy, targeted therapy or immunotherapy based on primary tumor histology according to national guidelines and at the investigators' discretion. The MTB also proposed possible modifications based on molecular profiling and expert opinion in the control group.

In the TT arm, patients received targeted therapy, immunotherapy or a combination that was selected according to their genomic profiles, as determined by the MTB. Combination therapies were proposed in cases where efficacy data for the combination were well established in different malignancies (for example, pertuzumab plus trastuzumab for *HER2*⁺ breast cancer and vemurafenib plus cobimetinib for *BRAF*-mutant melanoma) or when there was a biological rationale due to the presence of multiple actionable mutations. Such proposals were contingent upon the availability of published phase 1b/2 data supporting the combination in other tumor types. In cases involving immunotherapy, the combination of ipilimumab and nivolumab was usually preferred unless a patient's clinical condition or comorbidities suggested otherwise. The decision was made collaboratively between the MTB and the patient's treating clinician.

Patients were allowed to crossover to the TT arm upon disease progression. A substantial amendment to the protocol was implemented in October 2022, introducing additional treatments to the existing options, including entrectinib, alpelisib, pralsetinib and selipercatinib. Investigational therapies included erlotinib, pertuzumab, vemurafenib, trastuzumab emtansine, alectinib, vismodegib, cobimetinib, atezolizumab, trastuzumab, ipatasertib, entrectinib, everolimus, palbociclib, lapatinib, ipilimumab, nivolumab, brigatinib, ponatinib, itacitinib, pemigatinib, alpelisib, tepotinib, pralsetinib, talazoparib and selipercatinib. In cases where hTMB coexisted with a targetable mutation, a combination of immunotherapy and targeted therapy was generally preferred, provided that safety data were available. All treatments were administered according to the investigator's brochure or the relevant regulatory clinical protocol. Each targeted agent was associated with a specific pathway, signature or gene alteration (Extended Data Table 4).

Study endpoints

The primary endpoint of the study was the ORR, defined as the proportion of patients achieving a complete response or partial response, with respect to the total number of randomized patients. Secondary endpoints included PFS, defined as the time from randomization to disease progression or death, whichever occurred first; TTF, defined as the time from randomization until patient withdrawal for any reason, including disease progression or death; TTNT, defined as the time from randomization to the start of the next line of therapy; and OS, defined as the time from randomization to death from any cause. Additional planned secondary endpoints not reported in this paper include concordance between molecular profile on tumor tissue and ctDNA, quality of life measurements, immune fitness in the two treatment arms and the association between the molecular evaluation and gene expression profiling.

Assessment of treatment outcomes and safety

Tumor response was locally assessed according to RECIST version 1.1 and irRC. In patients who discontinued treatment for reasons other than disease progression or consent withdrawal, tumor assessments continued until documented progression. Survival was monitored until death, loss to follow-up, consent withdrawal or study termination. AEs were graded using CTCAE version 5.0, with the frequency and severity of AEs documented for each patient.

Data collection and trial oversight

Data were collected at the clinical sites and inserted into a web-based electronic case report form (eCRF) by investigators or designated personnel. The trial was overseen by an independent external data monitoring committee, which conducted periodic safety evaluations and reviewed efficacy during prespecified interim analyses. Per the study protocol, the data monitoring committee (DMC) conducted a prespecified interim analysis after 20% of the randomized patients ($n = 76$) had reached the final efficacy evaluation (day 308 or early termination), irrespective of treatment group assignment. The DMC reviewed these data and recommended proceeding with patient accrual as planned. As predefined, the stopping threshold for conditional power (CPt) was set at 5%. Comparisons were made between the observed TT group response rate at interim analysis and the projected end-of-study SoC group response rate (5%). At interim analysis, the TT group exhibited a 7.1% response rate. The calculated CPt for the projected SoC outcome, derived from the sample size assumptions, was 88.7%, demonstrating strong evidence to continue the trial and prompting a unanimous DMC recommendation to proceed. The authors confirm that the trial was conducted in compliance with the protocol, its amendments and the principles of Good Clinical Practice. All authors had access to the data used for manuscript preparation and contributed to its drafting, critical review or editing. The healthcare business of Merck KGaA, Darmstadt, Germany (CrossRef Funder ID: 10.13039/100009945), reviewed this manuscript for medical accuracy only before journal submission. The authors are fully responsible for the content of this paper, and the views and opinions described in the publication reflect solely those of the authors.

Sample size

We hypothesized that TT would yield higher ORR compared to SoC, specifically a 20% ORR for TT versus 5% for SoC. According to the site of primary tumor, four cohorts were defined as breast cancer (Stratum A), non-colorectal gastrointestinal cancers (Stratum B), NSCLC (Stratum C) and 'other malignancies' (Stratum D), with competitive enrollment across these strata. To detect a 15% difference between the two arms, assuming an α of 0.10 and a β of 0.20 (80% power) and employing a one-sided chi-square test, a total of 86 patients (43 in each arm of the four cohorts) were required. The four cohorts were evaluated as separate strata, resulting in a combined cohort size of

344 patients (86 patients per stratum). The overall ORR was assessed using a two-sided CMH test at a 5% significance level; an unstratified sensitivity analysis was also performed. Considering a dropout rate of 10%, 384 patients were required. Assuming actionable mutations in at least 30% of eligible patients, the planned screening sample size was 1,280. During the course of the study, the screening failure rate was higher than initially anticipated, leading to a total of 1,794 screened patients. Due to the extended screening periods required for genomic testing and the inherent unpredictability of patient eligibility based on mutational test results, 12 additional patients undergoing screening at the time the target of 384 randomized participants was reached were permitted by the steering committee to enroll, resulting in a final total of 400 patients.

Statistical analysis

All analyses were conducted using SAS software (v.9.4). The SAP, detailing the analyses to be executed, was finalized before database lock (SAP v.1.0). The randomization list was generated with a dedicated SAS program using the PROC PLAN procedure by an independent statistician. The randomization lists were incorporated into the eCRF system, which automatically managed the assignment of treatment arms. A soft lock of the database was performed on 31 July 2024. SAP v.2.0 was finalized on 30 January 2025, after the database freezing and data review meeting, and governs all analyses presented here. The full SAP is available upon reasonable request. Descriptive statistics were used to summarize the data, with mean, s.d., quartiles for continuous variables and frequency distributions for categorical variables. Differences in clinical characteristics were evaluated with *t*-test, binomial test, chi-square and CMH. The Breslow–Day test was used to evaluate the homogeneity of odds ratios. Box plots and tornado plots assessed the overall incidence of grade 3/4 AEs. Kaplan–Meier survival analysis, accompanied by the log-rank test, was employed to compare OS and PFS across arms and subgroups. To visualize clinical benefit duration (response and stable disease) across molecular subgroups—including high TMB and *PI3K/AKT*-altered and *ERBB2*-positive cohorts—swimmer plots were generated. HRs and 95% CIs were also calculated. Homogeneous subgroups of patients were evaluated in a prespecified exploratory analysis. Exploratory biomarker analyses, including assessments of hTMB (≥ 10 mutations per megabase), *BRAF* V600E mutations and *HER2* amplifications, were predefined in the SAP to evaluate treatment effect heterogeneity. These subgroups were selected based on emerging preclinical evidence during the trial's enrollment phase, with steering committee approval for post hoc stratification. All statistical analyses were conducted on the ITT population. R v.4.3.3 and RStudio were used to build toxicity and genomic alterations plots.

Reporting summary

Further information on research design is available in the Nature Portfolio Reporting Summary linked to this article.

Data availability

Individual deidentified participant data generated during the current study are available upon reasonable request from academic or qualified clinical researchers affiliated with recognized institutions, strictly for the purpose of conducting non-commercial, ethically approvable research aligned with the original scope of the trial. Applicants are required to submit a detailed research proposal, curriculum vitae and a declaration of non-conflict of interest. Requests must clearly describe the research objectives and methodology and must be reviewed and approved by the steering committee of the ROME trial during dedicated review sessions. Approval is granted based on scientific merit, data availability, intended data use and absence of overlapping research initiatives by the trial investigators. All approved requestors will be required to sign a data access agreement that restricts data use solely to the approved research project and prohibits any further distribution

or use. Data will be shared via a secure data-sharing platform within 4–8 weeks of approval, contingent upon data volume and complexity. Data requests will be considered within 12 months of manuscript publication.

The trial registration, study protocol and methodological details are publicly accessible through ClinicalTrials.gov (accession identifier [NCT04591431](https://clinicaltrials.gov/ct2/show/NCT04591431); <https://clinicaltrials.gov/ct2/show/NCT04591431>).

Additional publicly available datasets used in the analysis include the ClinVar database: freely accessible at <https://www.ncbi.nlm.nih.gov/clinvar/>; the OncoKB database: accessible at <https://www.oncokb.org/>; the COSMIC database: accessible at <https://cancer.sanger.ac.uk/cosmic>; and the ESMO ESCAT scale: accessible at <https://www.esmo.org/guidelines/esmo-scale-for-clinical-actionability-of-molecular-targets-escat>.

No other public repositories or datasets requiring accession codes were used in this study.

References

37. Eisenhauer, E. A. et al. New response evaluation criteria in solid tumours: revised RECIST guideline (version 1.1). *Eur. J. Cancer* **45**, 228–247 (2009).
38. Wolchok, J. D. et al. Guidelines for the evaluation of immune therapy activity in solid tumors: immune-related response criteria. *Clin. Cancer Res.* **15**, 7412–7420 (2009).
39. Wahl, R. L., Jacene, H., Kasamon, Y. & Lodge, M. A. From RECIST to PERCIST: evolving considerations for PET response criteria in solid tumors. *J. Nucl. Med.* **50**, 122S–150S (2009).

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Author contributions

Conceptualization: P.M., A.B. and S.S. Patient enrollment and investigation: G.C., M.B., S.L., S.S., L.F., V.G., U.D.G., P.A., M.A., C.C., P.C., E. Crimini, S.P., S.V., R. Bordonaro, S.B., G. Butturini, L.D.M., A.D.C., A.F., E.F., S.G., G.M., A.P., D.P., F.P., S.T., A.Z., D.M., F.C., F.M., G.T., A.B., A.S., V.A., D.Q., P.T., P.D., B.U., C.B., E.Z., C.P., A.T., L.B., E.M., C.M., A.R. and R. Berardi. MTB: P.M., G.C., M.B., S.S., P.A., G. Blandino, G.D., M.A., C.C., P.C., E. Capoluongo, F.C., B.C., G.G., U.M., F.M., M.N., G.P., L.S. and A.B. Methodology: L.S., M.C., P.M., A.B., S.S., M.B., P.A., G. Blandino, G.D., M.A., C.C., P.C., E. Capoluongo, F.C., B.C., G.G., U.M., F.M., M.N., G.P., N.M. and M.S. Software: M.C. and L.S. Formal analysis and

data curation: M.C. and L.S. Validation: P.M., A.B., S.S., L.S. and M.C. Writing—original draft preparation: P.M., S.S., E. Crimini, S.P., S.V. and A.B. Writing—review and editing: all authors. Visualization: P.M., S.S., E. Crimini, M.C. and A.B. All authors read and agreed to the published version of the manuscript.

Competing interests

P.M. had a consultant/advisory role for Bristol Myers Squibb, Roche Genentech, Merck Sharp & Dohme, Novartis, AMGEN, Merck Serono, Pierre Fabre and Incyte. He is a member of the advisory board of Drug-PIN. A.G. is the holder of patent PTC/IB2019/052310. G.C. had a role in advisory board for Roche, AstraZeneca, Daiichi Sankyo, Eli Lilly, Novartis, Pfizer, Gilead, Menarini, Exact Sciences, Bristol Myers Squibb and Merck. M.B. is employed in the Italian National Institute of Health (Istituto Superiore di Sanità) and is an unpaid member of the Technical and Scientific Committee of the Italian Medicine Agency (AIFA). S.L. was an invited speaker for Amgen, AstraZeneca, Bristol Myers Squibb, Incyte, GlaxoSmithKline, Eli Lilly, Merck Serono, Merck Sharp & Dohme, Pierre Fabre, Roche and Servier and had a role in advisory boards for Amgen, Astellas, AstraZeneca, Bayer, Bristol Myers Squibb, Daiichi Sankyo, GlaxoSmithKline, Incyte, Eli Lilly, Merck Serono, Merck Sharp & Dohme, Servier, Takeda, Rottapharm, BeiGene, Fosun Pharma and Nimbus Therapeutics. S.S. was an invited speaker for Pfizer, Eli Lilly, Novartis, Daiichi Sankyo, Gilead, Roche and AstraZeneca. L.F. reports speaking honoraria from Incyte, Bristol Myers Squibb and Eli Lilly. He also received institutional research funding from Merck Sharp & Dohme, Bristol Myers Squibb, AstraZeneca, Incyte, BeiGene, Astellas, Daiichi Sankyo and Roche. He had a role in advisory boards for Merck Sharp & Dohme, AstraZeneca, Incyte, Taiho, Servier, Daiichi Sankyo, Eli Lilly and Astellas. V.G. has a role in leadership for AstraZeneca, Daiichi Sankyo, Eli Lilly, Exact Sciences, Gilead, Merck Sharp & Dohme, Novartis, Pfizer, Olema Oncology, Pierre Fabre and Menarini Stemline. She had a role as a speaker for AstraZeneca, Daiichi Sankyo, Eli Lilly, Exact Sciences, Gilead, GlaxoSmithKline, Novartis, Roche, Zentiva and Menarini Stemline and provided expert testimony for Eli Lilly. U.D.G. received consultation fees from Amgen, Astellas, AstraZeneca, Bayer, Bristol Myers Squibb, Eisai, Ipsen, Johnson & Johnson Innovative Medicine (formerly Janssen), Merck KGaA, Merck Sharp & Dohme, Novartis and Pfizer and travel expenses for attending symposia from AstraZeneca, Ipsen and Pfizer. P.A., G.B., G.D., M.A., C.C., P.C., E. Crimini and M.C. declare no competing interests. S.P. had a role as consultant/advisor for AstraZeneca, Eli Lilly, Daiichi Sankyo, Novartis, Pfizer, Seagen, Sophos and Gilead and received travel and accommodation fees from Daiichi Sankyo, Pfizer, AstraZeneca, Eli Lilly, Menarini, Novartis and Gilead. S.V. had a role as a speaker for Novartis. R.B. declares no competing interests. S.B. received congress travel support from Merck Sharp & Dohme, Pfizer, Bayer and Ipsen and had a role in advisory boards and steering committees for Pfizer, Bristol Myers Squibb, Merck Sharp & Dohme, Astellas, Roche, Johnson & Johnson, Ipsen, Bayer, Novartis, Merck, AstraZeneca and Gilead. G.B. declares no competing interests. L.D.M. received personal grant for advisory/consultant/speaker activities from Agendia, AstraZeneca, Daiichi Sankyo, Eli Lilly, Eisai, Exact Sciences, Gilead, GlaxoSmithKline, Ipsen, Roche, Seagen, Menarini, Stemline, Merck Sharp & Dohme, Novartis, Olema, Pierre Fabre and Pfizer. A.D.C. declares no competing interests. A.F. received financial fees from Roche, Menarini, Pfizer, Eli Lilly, Novartis, Daiichi Sankyo, AstraZeneca and Gentili. E.F., S.G., G.M., A.P. and D.P. declare no competing interests. F.P. received honoraria for advisory boards, activities as a speaker, travel grants and research grants from Amgen, AstraZeneca, Daiichi Sankyo, Celgene, Eisai, Eli Lilly, Exact Sciences, Gilead, Ipsen, Italfarmaco, Menarini, Merck Sharp & Dohme, Novartis, Pierre Fabre, Pfizer, Roche, Seagen, Takeda and Viatrix. He received research funding from AstraZeneca, Eisai and Roche. S.T. declares no competing interests. A.Z. received

honoraria for consultancy and advisory board from Roche, Novartis, Eli Lilly, AstraZeneca, Pfizer, Exact Sciences, Merck, Daiichi Sankyo, Gilead, Seagen, Menarini and Stemline. D.M. had a role in advisory boards for Roche, Merck Sharp & Dohme, Merck and AstraZeneca and received fees for travel and meetings from Pierre Fabre and Amgen. F.C. received fees for membership of an advisory board or lectures from Roche, AstraZeneca, Bristol Myers Squibb, Pfizer, Takeda, Eli Lilly, Bayer, Amgen, Sanofi, Pharmamar, Novocure, Mirati, Galecto, OSE, Illumina, Thermo Fisher Scientific, BeiGene and Merck Sharp & Dohme. B.C. had a role in advisory boards for Merck Sharp & Dohme and Daiichi Sankyo. G.G. declares no competing interests. U.M. has received personal fees (as a consultant and/or speaker bureau) from Boehringer Ingelheim, Roche, Merck Sharp & Dohme, Amgen, Thermo Fisher Scientific, Eli Lilly, Diaceutics, GlaxoSmithKline, Merck, AstraZeneca, Janssen, Diotech, Novartis and Heder. F.M. and M.N. declare no competing interests. G.P. received financial fees from Illumina, Roche, Eli Lilly, AstraZeneca, Exact Sciences and ADS Biotech. M.S. and L.S. declare no competing interests. G.T. had a role in advisory boards for Molteni, Novartis, PharmaMar and Merck Sharp & Dohme. N.M. received advisory board consultation fees from Roche, Daiichi Sankyo, Pfizer and AstraZeneca. A.B. had a role in advisory boards and steering committees and has been an invited speaker for Novartis, Roche, Eli Lilly, Pfizer, AstraZeneca, Daiichi Sankyo, Gilead, Merck Sharp & Dohme, Bristol Myers Squibb and Gentili. A.S., V.A., D.Q., P.T., P.D., B.U., C.B. and E.Z. declare no competing interests. C.P. received personal fees for advisory role, speaker engagements and travel accommodation expenses from Amgen, Astellas, AstraZeneca, Bayer, Bristol Myers Squibb, Celgene, Daiichi Sankyo, Eisai, Ipsen, Janssen, Incyte, Merck Serono, Merck Sharp & Dohme, Novartis, Roche, Sandoz, Sanofi and Servier. A.T., L.B., E.M., C.M. and A.R. declare no competing interests. R. Berardi received consulting fees and/or donations to institution from AstraZeneca, Pfizer, Roche, Astellas, Eisai, Merck Sharp & Dohme, Menarini, Gilead, Pierre Fabre, Bristol Myers Squibb, Seagen, Incyte, Eli Lilly, Bayer and Daiichi Sankyo. G.L. received advisory/consulting fees from AbbVie, Bayer, Novartis, Orbus, Celgene, GlaxoSmithKline, Health4u, Braun Melsungen, Janssen, BioRegio-Stern, Servier, Novocure and TME Pharma and travel fees from Roche, Bayer and Servier. A. Piancastelli declares no competing interests. G. Masi received fees for advisory boards from AstraZeneca, Bayer, Ipsen, Merck Sharp & Dohme, Eisai, Roche and Sirtex; speaking honoraria from Amgen, AstraZeneca, Roche, Merck Sharp & Dohme, Eisai, Terumo and Sirtex; travel grants from AstraZeneca, Bayer, Ipsen, Merck Sharp & Dohme, Eisai and Roche; and research grants from Roche, Terumo and Sirtex. L.B., V.V. and F. Mannozi declare no competing interests.

Additional information

Extended data is available for this paper at <https://doi.org/10.1038/s41591-025-03918-x>.

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Correspondence and requests for materials should be addressed to Simone Scagnoli.

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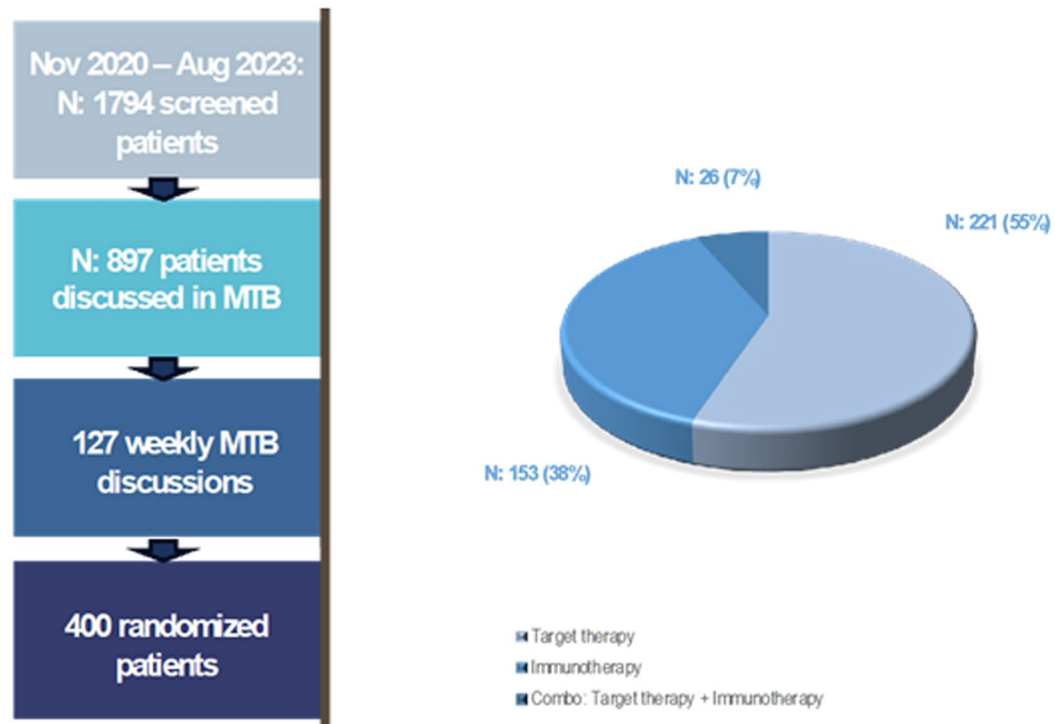
Alteration Type

- Amplification
- Deletion
- Mutation
- Mutation and Amplification
- Rearrangement and Amplification
- Rearrangement/Fusion
- Signature

Gene(s)	Count	Alteration Type(s)
TMB-H	135	Signature
MSI-H	18	Signature
RET	2	Rearrangement/Fusion
MAP2K1/2	2	Mutation
GNA11	2	Mutation
NRAS	5	Mutation
NF1	4	Mutation
KRAS	4	Mutation
BRAF	14	Mutation
RAS/RAF/MAPK	-	Mutation
PTCH1	2	Mutation
PIK3CA/AKT/PEN	-	Mutation
PIK3R1	3	Mutation
AKT2	4	Mutation
AKT1	3	Mutation
PTEN	40	Deletion
PIK3CA	49	Mutation
STK11	7	Mutation
FBXW7	7	Mutation
Other	-	Mutation
JAK/STAT/JAK2	-	Mutation
MET	1	Mutation
HRD	3	Mutation
BRCA2	2	Mutation
BRCA1	2	Mutation
ATM	1	Mutation
PALB2	1	Mutation
ErbB	7	Amplification
ERBB3	7	Amplification
CDKN2A	4	Amplification
CDK4	7	Amplification
Cell cycle	7	Amplification
FGFR	12	Amplification
FGFR1	12	Amplification
FGFR2	12	Amplification
FGFR3	7	Amplification
FGF10	1	Amplification
FGF3	1	Amplification
ALK/ROS1	1	Rearrangement and Amplification
ROS1	1	Rearrangement and Amplification
ERBB2	57	Amplification

Alteration group	Therapy assigned	n
Signature n = 153	Ipilimumab + Nivolumab	148
	Nivolumab	5
PIK3CA/AKT/PTEN n = 99	Ipatasertib	66
	Atezolizumab + Ipatasertib	24
	Alpelisib	6
	Everolimus	3
Erb n = 58	TDM1	31
	Pertuzumab + Trastuzumab	12
	Trastuzumab + Lapatinib	11
	Pertuzumab + TDM1	2
	TDM1 + Atezolizumab	1
	Trastuzumab + Everolimus	1
FGFR n = 33	Pemigatinib	33
RAS/RAF/MAPK n = 31	Vemurafenib + Cobimetinib	16
	Cobimetinib	14
	Cobimetinib + Atezolizumab	1
HRD n = 7	Talazoparib	7
ALK/ROS1 n = 6	Alectinib	4
	Entrectinib	2
Cell cycle n = 5	Palbociclib	5
PTCH1 n = 2	Vismodegib	2
RET n = 2	Pralsetinib	1
	Selpercatinib	1
Other n = 2	Everolimus	2
JAK/STAT n = 1	Itacitinib	1
MET n = 1	Tepotinib	1

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Extended Data Fig. 2 | Molecular tumor board activity and treatment outcomes in the intention-to-treat (ITT) population. MTB outcomes. Molecular tumor board activity and treatment allocation in the intention-to-treat (ITT) population.

Grade ≥ 3 Adverse events

A

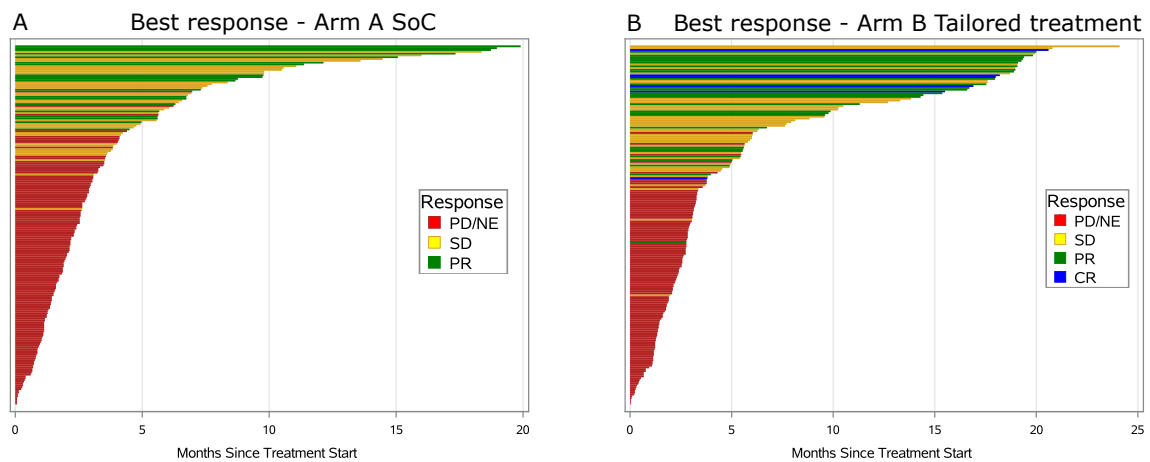
Organ or system (CTCAE v5)	Adverse event	Overall 400 n (%)	SOC 200 n (%)	TT 200 n (%)
Investigations	Neutrophil count decreased	21 (5.2%)	16 (8%)	5 (2.5%)
General and administration site	Fatigue	18 (4.5%)	12 (6%)	6 (3%)
Gastrointestinal	Diarrhoea	14 (3.5%)	3 (1.5%)	11 (5.5%)
Blood and lymphatic system	Anaemia	13 (3.2%)	10 (5%)	3 (1.5%)
Investigations	Blood Bilirubin increased	11 (2.8%)	9 (4.5%)	2 (1%)
Investigations	Transaminases increased	11 (2.8%)	4 (2%)	7 (3.5%)
Infections and infestations	Sepsis	9 (2.2%)	8 (4%)	1 (0.5%)
General and administration site	Pain	7 (1.8%)	3 (1.5%)	4 (2%)
Gastrointestinal	Abdominal pain	6 (1.5%)	3 (1.5%)	3 (1.5%)
Infections and infestations	Lung infection	6 (1.5%)	4 (2%)	2 (1%)
Respiratory, thoracic and mediastinal	Dyspnea	6 (1.5%)	5 (2.5%)	1 (0.5%)
Gastrointestinal	Intestinal obstruction	5 (1.2%)	3 (1.5%)	2 (1%)
General and administration site	Malaise	5 (1.2%)	1 (0.5%)	4 (2%)
Metabolism and nutrition	Hyperglycemia	5 (1.2%)	2 (1%)	3 (1.5%)
Skin and subcutaneous tissue	Rash	5 (1.2%)	3 (1.5%)	2 (1%)
Vascular	Hypertension	5 (1.2%)	3 (1.5%)	2 (1%)

B

Organ or system (CTCAE v5)	Adverse event	Arm	Drugs	Treatment-related	Notes
Infections and infestations	Lung infection	SOC	FOLFIRI	No	COVID-19
Infections and infestations	Sepsis	SOC	FOLFIRI	No	
Infections and infestations	Sepsis	TT	Nivolumab + Ipilimumab	Yes	
Respiratory, thoracic and mediastinal	Pneumonitis/ILD	SOC	Paclitaxel	Yes	
Vascular disorders	Ischemia cerebrovascular	TT	Nivolumab	No	
Vascular disorders	Thromboembolic event	TT	Pralsetinib	No	Pulmonary thromboembolism
Cardiac disorders	Cardiac arrest	SOC	FOLFIRI	No	
Nervous system disorders	Encephalitis	TT	Nivolumab + Ipilimumab	Yes	

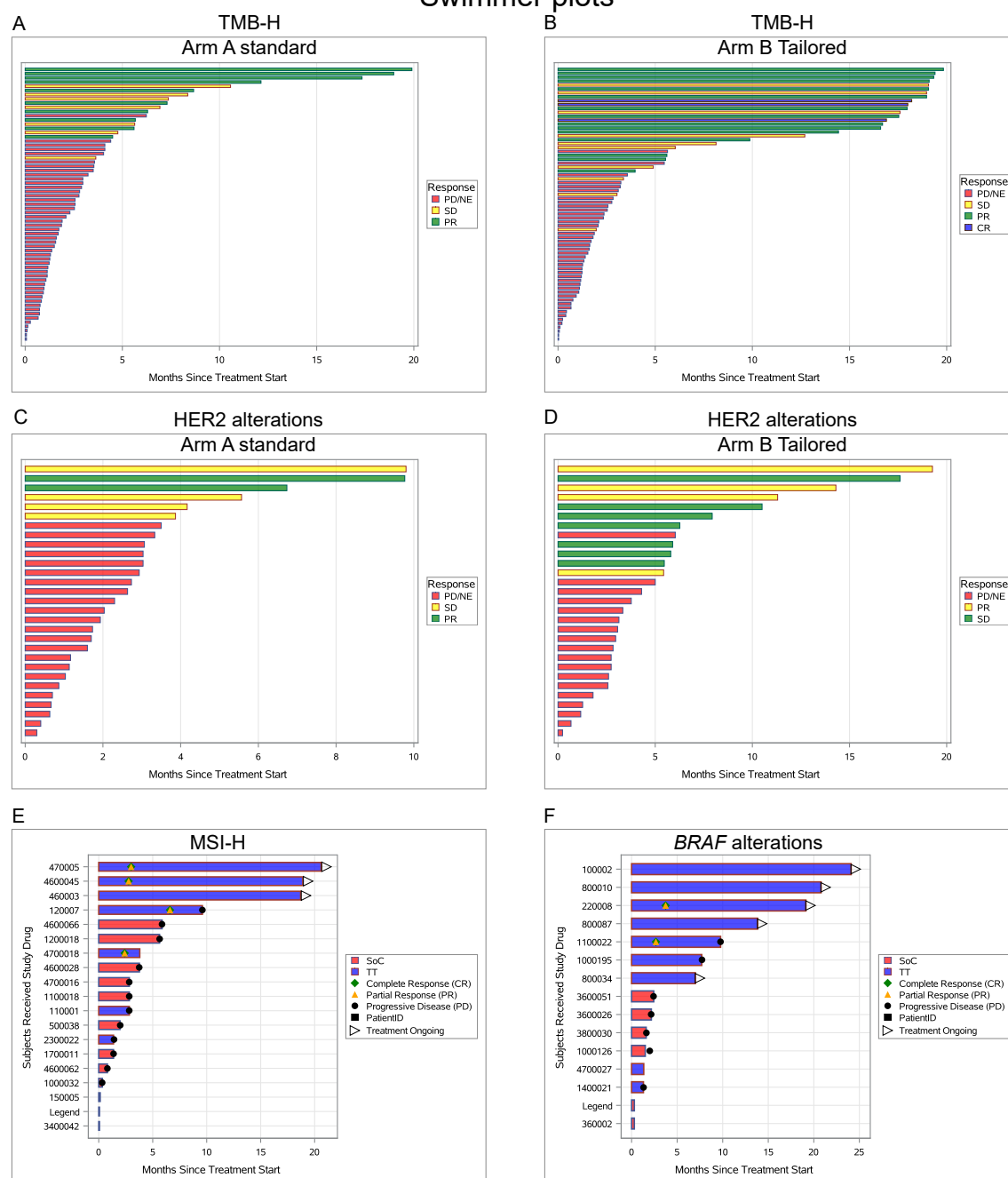
Extended Data Fig. 3 | Grade ≥ 3 Adverse events. **a)** Grade ≥ 3 adverse events that occurred in at least 5 patients. **b)** Grade 5 adverse events. Each line reports a different event. SoC: standard of care, TT: tailored treatment; CTCAE: Common Terminology Criteria for Adverse Events.

Swimmer plots in intention to treat population



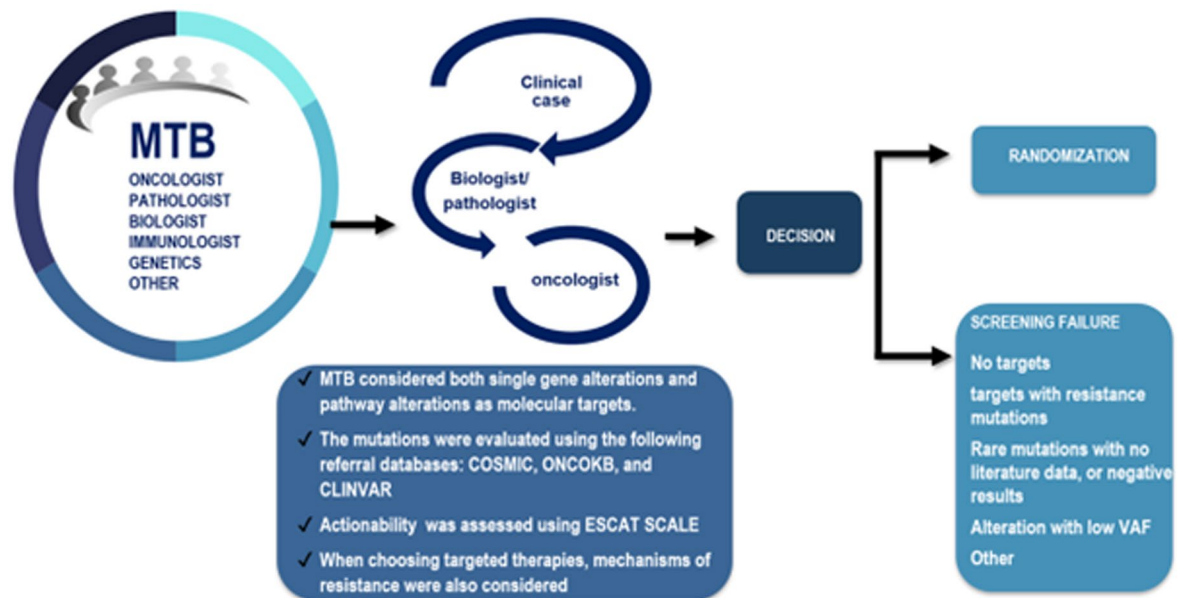
Extended Data Fig. 4 | Swimmer plots for SoC and TT (ITT population). a) SOC; b) TT. TT: tailored treatment; SOC: standard of care; PD: Progressive disease; NE: Not evaluable; SD: Stable disease; PR: Partial response; CR: Complete response; ITT: intention-to-treat.

Exploratory subgroup analysis Swimmer plots



Extended Data Fig. 5 | Swimmer plots for SoC and TT in exploratory hTMB, MSI, BRAF and HER2 subgroups. a) hTMB SOC; b) hTMB TT; c) HER2 alterations SOC; d) HER2 alterations TT; e) MSI-H; f) BRAF alterations. TT: tailored

treatment; SOC: standard of care; PD: Progressive disease; NE: Not evaluable; SD: Stable disease; PR: Partial response; CR: Complete response; hTMB: High tumor mutational burden; MSI-H: microsatellite instability high.



Extended Data Fig. 6 | MTB structure and activities. The Molecular Tumor Board (MTB)'s organizational structure and core activities illustrate its workflow and multidisciplinary collaboration for precision oncology decision-making.

Extended Data Table 1 | ORR in ITT population

Strata	Response	SoC N=200 (%)	TT N=200 (%)
Stratum A: Breast	CR/PR	7 (35.0%)	4 (20.0%)
	SD/NE/PD	13 (65.0%)	16 (80.0%)
Stratum B: Gastrointestinal Cancer	CR/PR	5 (9.1%)	8 (14.8%)
	SD/NE/PD	50 (90.9%)	46 (85.2%)
Stratum C: Non-small cell Lung Cancer	CR/PR	1 (6.7%)	2 (12.5%)
	SD/NE/PD	14 (93.3%)	14 (87.5%)
Stratum D: Other	CR/PR	7 (6.4%)	21 (19.1%)
	SD/NE/PD	103 (93.6%)	89 (80.9%)
p-value		0.0285	

ORR in the ITT population, summarizing the proportion of patients achieving a predefined clinical response to the treatment. CR, complete response; PD, progressive disease; PR, partial response; SD, stable disease.

Extended Data Table 2 | ORR as per strata

Best Overall Response	Overall N=400 (%)	SoC N=200 (%)	TT N=200 (%)
CR	6 (1.5%)	0 (0.0)	6 (3.0%)
PR	49 (12.3%)	20 (10.0%)	29 (14.5%)
SD	74 (18.0%)	35 (17.5%)	39 (19.5%)
PD	271 (67.8%)	145 (72.5%)	126 (63.0%)
Overall Response Rate	13.8% [CI 95% 10.5-17.5]	10% [CI 95% 6.2-15.0]	17.5% [CI 95% 12.5-23.5]
p-value	0.0294		

ORR analyzed according to stratified subgroups. CR, complete response; PD, progressive disease; PR, partial response; SD, stable disease.

Extended Data Table 3 | Treatment response in exploratory hTMB, MSI, BRAF and HER2 subgroups

Best Overall Response	Overall	SoC	TT
hTMB subgroup	N=135 (%)	N=65 (%)	N=70 (%)
CR	3 (2.2)	0 (0.0)	3 (4.3)
PR	25 (18.5)	10 (15.4)	15 (21.4)
SD	17 (12.6)	7 (10.8)	10 (14.3)
PD	90 (66.7)	48 (73.9)	42 (60.0)
ORR% (95% CI)	20.7 (14.3-28.6)	15.4 (7.6-26.5)	25.7 (16.0-37.6)
p-value	0.1391		
MSI-H subgroup	N=18 (%)	N=11 (%)	N=7 (%)
CR	2 (11.1)	0 (0.0)	2 (28.6)
PR	2 (11.1)	0 (0.0)	2 (28.6)
SD	3 (16.7)	2 (18.2)	1 (14.2)
PD	11 (61.1)	9 (81.8)	2 (28.6)
ORR% (95% CI)	22.2 (6.4-47.6)	0 (0.0)	57.1 (18.4-90.1)
p-value	0.0114		
BRAF subgroup	N=14 (%)	N=5 (%)	N=9 (%)
CR	0 (0.0)	0 (0.0)	0 (0.0)
PR	2 (14.3)	0 (0.0)	2 (22.2)
SD	5 (35.7)	0 (0.0)	5 (55.6)
PD	7 (50.0)	5 (100.0)	2 (22.2)
ORR% (95% CI)	14.3 (1.8-42.8)	0 (0.0)	22.2 (2.8-60.0)
p-value	0.5054		
HER2 subgroup	N=58 (%)	N=29 (%)	N= 29 (%)
CR	0 (0.0)	0 (0.0)	0 (0.0)
PR	6 (10.3)	2 (6.9)	4 (13.8)
SD	11 (19.0)	4 (13.8)	7 (24.1)
PD	41 (70.7)	23 (79.3)	18 (62.1)
ORR% (95% CI)	810.3 (3.9-21.2)	6.9 (0.9-22.8)	13.8 (3.9-31.7)
p-value	0.6701		

Treatment response observed in exploratory subgroups, including hTMB, MSI, *BRAF* mutations and *HER2* alterations. CR, complete response; PD, progressive disease; PR, partial response; SD, stable disease.

Extended Data Table 4 | Available targeted treatments and pathways

Target pathways	Targeted Therapy
EGFR mutations	Erlotinib
ERBB2 amplifications/mutations	Trastuzumab, pertuzumab, TDM1, lapatinib
mTOR mutations, AKT mutations	Everolimus
BRAF mutations	Vemurafenib, cobimetinib
ALK translocations/RET alterations	Alectinib, brigatinib
RET	Pralsetinib*, selpercatinib*
CDK4/6I, CDKN2A/B	Palbociclib
Bcr-abl	Ponatinib
SMOP/PTCH1	Vismodegib
JAK mutations	Itacitinib
FGFR1/2/3 alterations	Pemigatinib
PI3K/AKT, PTEN alterations	Ipatasertib, alpelisib*
NTRK1/2/3 fusions, ROS1	Entrectinib*
METex14, MET alterations	Tepotinib
BRCA 1/2, HRD	Talazoparib
Biomarkers	Immunotherapy
MSI, high TMB**	Nivolumab, ipilimumab, atezolizumab

Overview of available targeted treatments and their associated pathways. * Drugs included in the V4 protocol amendment in October 2022. ** hTMB defined as ≥ 10 mutations per megabase in solid and/or liquid NGS.

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ClinVar database: freely accessible at <https://www.ncbi.nlm.nih.gov/clinvar/>

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Participants in the ROME trial were recruited prospectively from specialized oncology centers. Eligible consecutive patients were identified and invited to participate based on clear inclusion and exclusion criteria detailed in the published protocol (NCT04591431). Recruitment involved direct clinician referral and systematic screening of eligible patients from participating institutions. Considering the broad inclusion criteria, the agnostic purpose of the trial, and that all the procedures and the NGS testing were included in the trial procedures, no selection bias or self-selection bias is expected.

Ethics oversight

The study was approved by the Institutional Ethical Committee of the coordinating center (Sapienza no. rif. C.E. 5575 - 02/2020) and by the Ethical Committee of each participating center. The investigational sites that approved the study protocol were: AOU Policlinico Umberto I, Rome; European Institute of Oncology (IEO), IRCCS; Milan; Veneto Institute of Oncology IOV-IRCCS, Padua; Division of Medical Oncology, Pisa University Hospital; IRCCS Istituto Romagnolo per lo Studio dei Tumori (IRST) "Dino Amadori", Meldola; Istituto Nazionale Tumori IRCCS Fondazione "G. Pascale", Naples; IRCCS Istituto Nazionale Tumori Regina Elena (IRE), Rome; IRCCS - Istituto di Candiolo, Candiolo; Medical Oncology Unit - ARNAS Garibaldi Catania; Azienda Ospedaliera Santa Maria di Terni, Terni; Pederzoli Hospital, Peschiera del Garda; IRCCS Ospedale Policlinico San Martino, Genoa; Ospedali Galliera, Genoa; Central Hospital of Belcolle, Viterbo; IRCCS Sacro Cuore Don Calabria Hospital, Negrar di Valpolicella; Santa Maria della Misericordia Hospital, Perugia; Nuovo Ospedale di Prato-Santo Stefano, Azienda USL Toscana Centro, Prato; CRO Aviano, National Cancer Institute, IRCCS, Aviano; Santa Maria delle Croci Hospital, AUSL Romagna, Ravenna; Oncology Unit ASST Papa Giovanni XXIII, Bergamo; Ordine Mauriziano Hospital, Turin; AOU Policlinico S. Andrea, Rome; Azienda Ospedaliera Universitaria Federico II, Napoli; Fondazione IRCCS Istituto Tumori di Milano; Fondazione Policlinico Universitario Campus Bio-Medico, Rome; A.O. Papardo-Messina; IRCCS Istituto Tumori "Giovanni Paolo II", Bari; Policlinico universitario "Mater Domini", Catanzaro; Centro Oncologico San Leopoldo Mandic, Isola Tiberina Gemelli isola; Rome; University Hospital, Ferrara; Misericordia Hospital, Grosseto; AUSL Piacenza Guglielmo da Saliceto Hospital, Piacenza; Comprehensive Cancer Centre, AUSL-IRCCS di Reggio Emilia, Reggio Emilia; Humanitas Gradenigo, Torino; Foundation IRCCS Casa Sollievo della Sofferenza, San Giovanni Rotondo; Ramazzini Hospital, Azienda Unità Sanitaria Locale Modena (AUSL), Carpi; AOUP "Paolo Giaccone", Palermo; AOU delle Marche, Ancona.

The competent authority Agenzia Italiana del Farmaco (AIFA) authorized the trial on 08/July/2020 (AIFA/SC/P/76132). The trial was registered in ClinicalTrials.gov with ClinicalTrials.gov ID NCT04591431. EudraCT number: 2018-002190-21. The trial adhered to the principles of the Declaration of Helsinki regarding research involving human subjects. 41 centers received ethical approval and participated in the study enrollment. All patients signed the specifically conceived informed consent form.

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Sample size	We hypothesized that tailored therapy (TT) would yield higher ORR compared to SoC, specifically a 20% ORR for TT versus 5% for SoC. According to the site of primary tumor, four cohorts were defined as breast cancer (stratum A), non-colorectal gastrointestinal cancers (stratum B), NSCLC (stratum C), and other malignancies (stratum D), with competitive enrollment across these strata. To detect a 15% difference between the two arms, assuming an alpha of 0.10, a beta of 0.20 (80% power), and employing a one-sided Chi-square test, a total of 86 patients (43 in each arm of the four cohorts) were required.
Data exclusions	no data were excluded. CONSORT diagram is included in the manuscript
Replication	Next-generation sequencing (NGS) tests were performed centrally by Foundation One Inc., utilizing standardized protocols and comprehensive quality control measures provided by the company. Each test was conducted only once per tissue (paraffin-embedded slides) and blood samples. Residual tissue samples, if any, were returned to the recruiting centers, which managed their return to local pathology archives to remain available for the patient. Blood samples could only be used once, making the test inherently non-repeatable.
Randomization	patients were randomized 1:1 to receive Tailored Treatment (TT) or Standard of Care (SoC). SAP 2.0 is provided
Blinding	open randomization

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Study protocol	included in the supplementary materials
Data collection	Data were collected from every site investigators in a specific eCRF. Recruitment was active from 13th October 2020 to 19th July 2023. Data collection ended on 30/Jan/2025 due to database lock. 1794 patients were screened. 400 were randomized.
Outcomes	The primary endpoint of the study was the overall response rate (ORR), defined as the proportion of patients achieving a complete response (CR) or partial response (PR), respect to the total number of randomized patients. Secondary endpoints included progression-free survival (PFS), defined as the time from randomization to disease progression or death, whichever occurred first; time to treatment failure (TTF), defined as the time from randomization until patient withdrawal for any reason including disease progression or death; time to next treatment (TTNT), defined as the time from randomization to the start of the next line of therapy; overall survival (OS), defined as the time from randomization to death from any cause and safety. Tumor response was locally assessed according to the RECIST version 1.1 and irRC criteria. Adverse events (AEs) were graded using the Common Terminology Criteria for Adverse Events (CTCAE) version 5.0. Additional planned secondary endpoints not reported in this manuscript are: concordance between molecular profile on tumor tissue and ctDNA, Quality of Life (QoLs), the immune fitness in the two treatment arms, and the association between the molecular evaluation and gene expression profiling.

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