

The Lipidome: A Case Study of Treatment-Resistance in Clear Cell Renal Cell Carcinoma using Patient Derived Organoids

Author: Sultan Chatila Supervisor: Prof. David Harrison



Summary

Purpose: To investigate the effects of three anti-cancer drugs on the lipidome of clear cell renal cell carcinoma (ccRCC) and identify lipids associated with treatment resistance and/or susceptibility

Rationale: ccRCC is extremely resistant to treatment; 1 in 2 patients will not survive 5 years post-diagnosis. Lipids are known to contribute to treatment resistance in other cancers yet their role in ccRCC remains poorly understood.

Method: Patient-derived organoids were treated with three anti-cancer drugs. The lipidomes of treated and untreated (control) PDOs were identified and compared

Main finding: Glycerophospholipids may contribute to treatment resistance in ccRCC.

Significance: Identified glycerophospholipids as potential targets for new generation anti-cancer drugs, which could bypass ccRCC resistance thus improving patient survival in the future.

Introduction

Background:

- Clear Cell Renal Cell Carcinoma (ccRCC) is the predominant type of Kidney Cancer, the 8th most common cancer globally
- Characterised by high lipid abundance
- Lipids contribute to pathways in various cancers that **protect tumour cells from treatment**¹
- However, lipid changes associated with treatment resistance in ccRCC remain **poorly characterized**^{2,3}

Objectives:

- Uncover effects of NUC-7738 (novel anti-cancer drug), Nivolumab and Ipilimumab on the lipidome (lipid composition) of ccRCC
- Identify lipids associated with treatment resistance and/or susceptibility

Methods

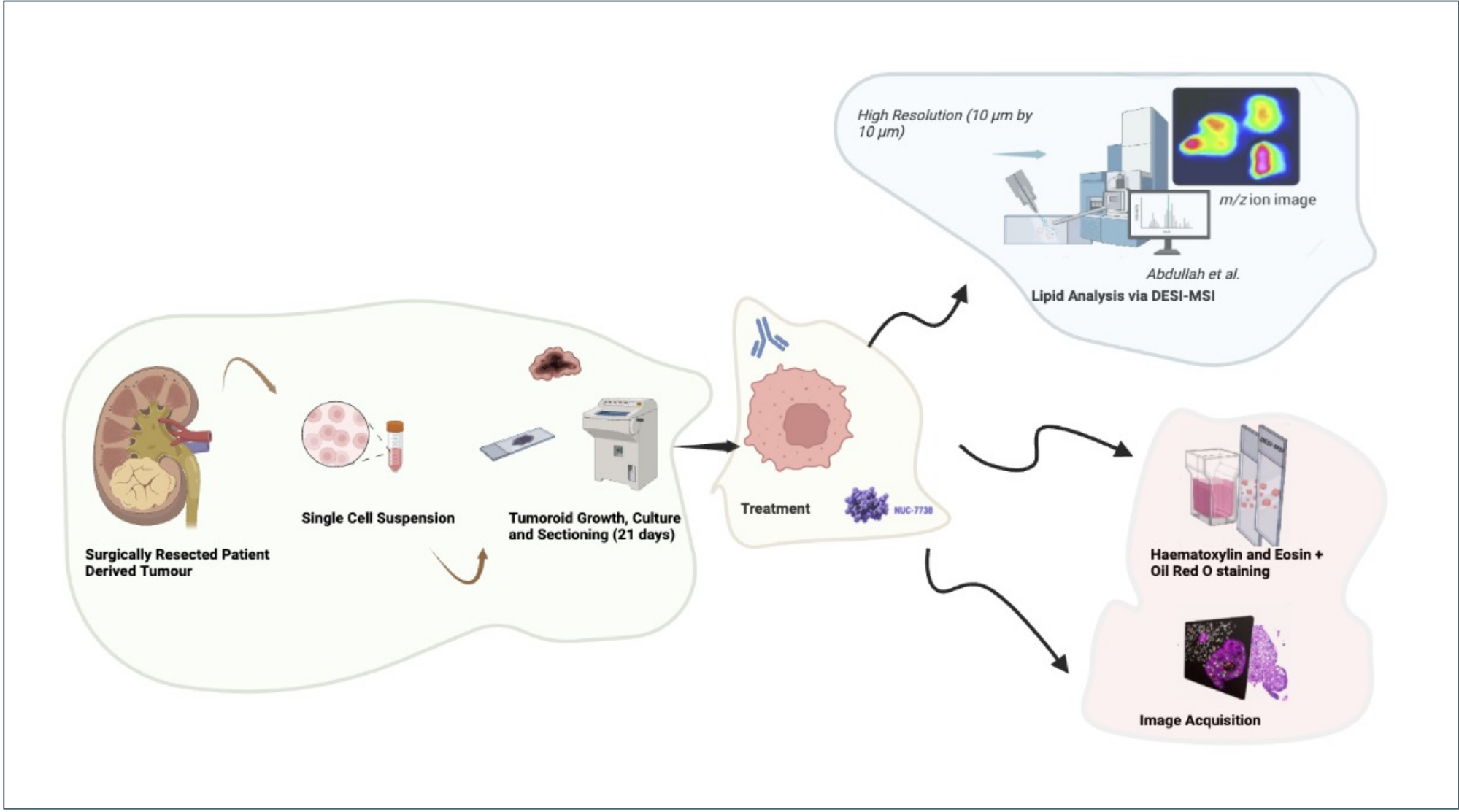


Fig. 1: Diagram outlining the harvesting, preparation, treatment and subsequent analysis of ccRCC organoids

- ccRCC cells taken from patient tumours (Fig. 2)
- Isolated patient ccRCC cells cultured to create **Patient Derived Organoids** (PDOs) (Fig. 3)
 - PDOs are 3D models of cancer cells which imitate the **structure and behaviour** of cancer in the human body
- Lipidome of each PDO was captured using **mass spectrometry** (Fig 4)

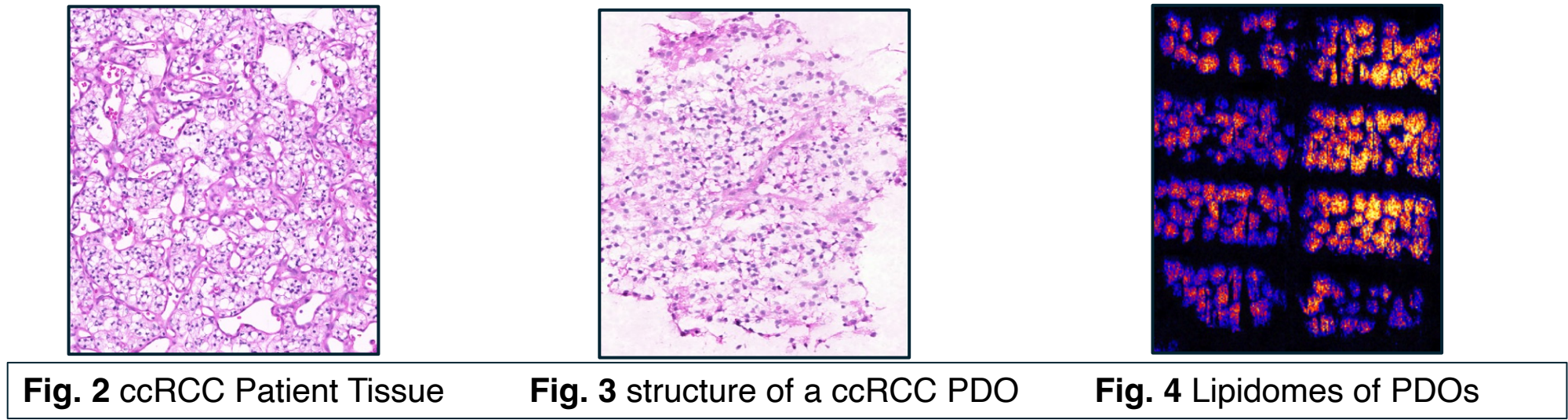


Fig. 2 ccRCC Patient Tissue Fig. 3 structure of a ccRCC PDO Fig. 4 Lipidomes of PDOs

Results

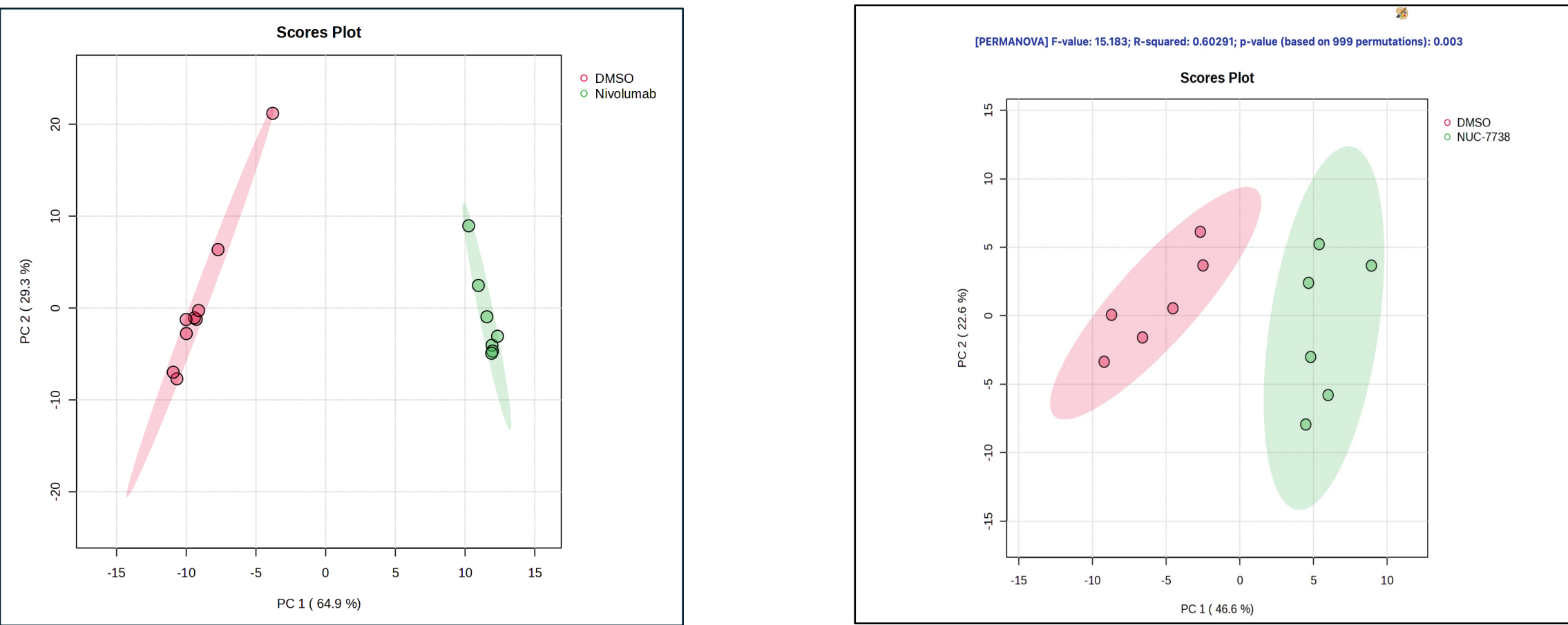


Fig. 5a (left) and 5b (right): Principal Component Analysis (PCA) , a pattern-finding statistical tool. PCA compares the lipid profiles of control (red) and treated (green) samples. Each circle represents **one sample**. (Images above highlight examples of PCA for two of the tested drugs)

- All tested anti-cancer drugs induced **changes in the lipidome** of ccRCC PDOs compared to control

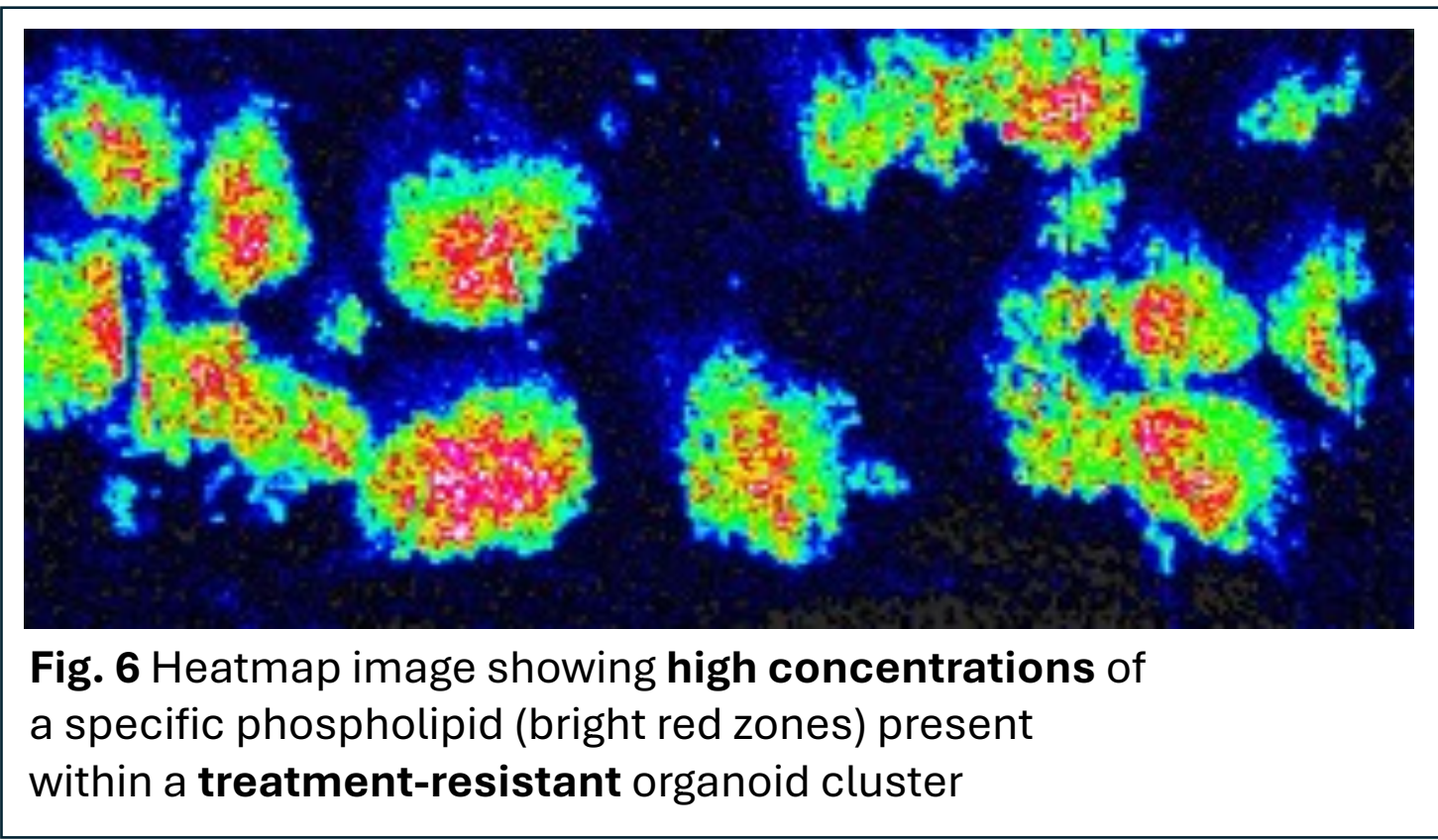


Fig. 6 Heatmap image showing **high concentrations** of a specific phospholipid (bright red zones) present within a **treatment-resistant** organoid cluster

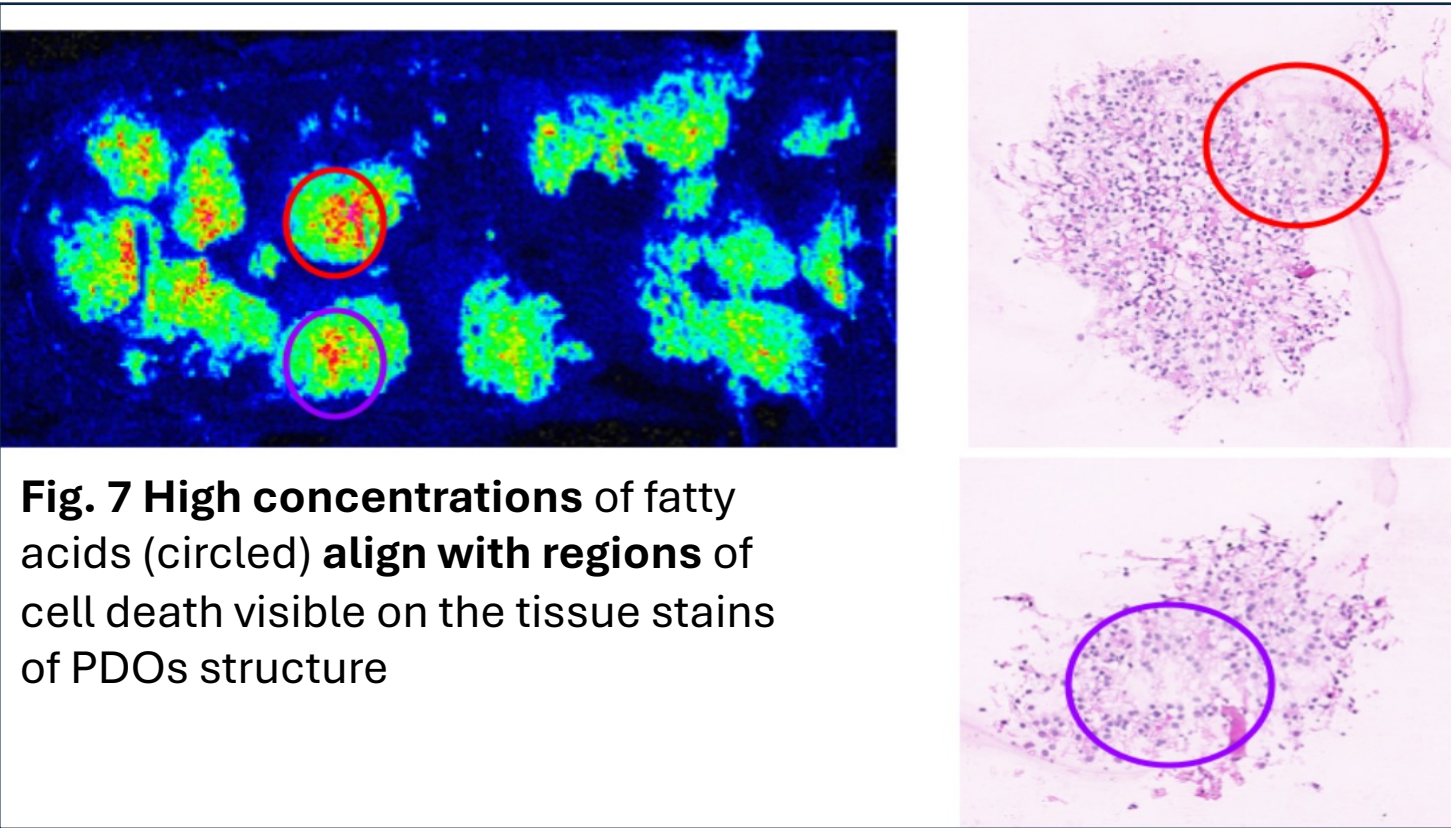


Fig. 7 High concentrations of fatty acids (circled) align with regions of cell death visible on the tissue stains of PDOs structure

- Glycerophospholipids** were present in higher abundance within PDOs that did not respond to treatment

- Fatty acids** were present in higher abundance within treatment responsive PDOs
- Fatty acids were associated with areas of cell death

Discussion

- The findings suggest that glycerophospholipids may play a role in treatment resistance and that fatty acids might be associated with treatment susceptibility
- Glycerophospholipids influence cellular signalling, cancer progression, and immune-cell function which **contribute to treatment resistance** in other cancers^{4,5}
- Drugs** targeting glycerophospholipids given in tandem with current treatments can potentially help **counter resistance** in ccRCC
- Fatty acids are known to drive and accumulate as byproducts of cell death⁶
- Therefore, the confinement of fatty acids to areas of cell death in our samples can confound their influence in ccRCC treatment susceptibility
- Future studies should **expand the sample size** beyond N=1 to improve generalizability
- Use of PDOs is a **strength** as it allows accurate bridging of relevant scientific findings to patients

Acknowledgements:

I would like to express my deepest gratitude to Lord Laidlaw and the Laidlaw Scholars Foundation for providing me with funds and preparation to carry out this project. I would also like to thank my supervisor Prof. David Harrison and his research team, namely Greice Zickuhr for to take part in your projects. You have been amazing mentors and supporters and I am very lucky to have had the chance to work with and learn from you.

References:

- Abdullah, H., et al. (2025). Kidney tumour characterisation by spatial mass spectrometry with same-section multiplex immunofluorescence uncovers tumour microenvironment lipid signatures associated with aggressive tumour phenotypes. *Npj Imaging*, 3(1). <https://doi.org/10.1038/s44303-025-00106-x>
- Li, W., Wang, X., Zhang, X., Gong, P., Ding, D., Wang, N., & Wang, Z. (2021). Revealing potential lipid biomarkers in clear cell renal cell carcinoma using targeted quantitative lipidomics. *Lipids in Health and Disease*, 20(1), 160. PubMed. <https://doi.org/10.1186/s12944-021-01572-z>
- Bobulescu, I. A., Pop, L. M., Mani, C., Turner, K., Rivera, C., Khatoon, S., Kairamkonda, S., Hannan, R., & Palle, K. (2021). Renal lipid metabolism abnormalities in obesity and clear cell renal cell carcinoma. *Metabolites*, 11(9), 608. PubMed Central. <https://doi.org/10.3390/metabo11090608>
- Preetam, S., et al. (2024) *Phosphatidylserine: paving the way for a new era in cancer therapies*. Rsc.Org. <https://pubs.rsc.org/ma/article/5/2/18384/853958/Phosphatidylserine-paving-the-way-for-a-new-era-in>
- Zhao, H., et al. (2025). Phospholipid metabolism and drug resistance in cancer. *Life Sciences*, 372, 123626. PubMed. <https://doi.org/10.1016/j.lfs.2025.123626>
- Fatty acids and their lipid mediators in the induction of cellular apoptosis in cancer cells. (2022). *Prostaglandins & Other Lipid Mediators*, 160, 106637. <https://doi.org/10.1016/j.prostaglandins.2022.106637>



Please Scan for full report